ARTICLE

Circulating Carotenoids and Risk of Breast Cancer: Pooled Analysis of Eight Prospective Studies

A. Heather Eliassen, Sara J. Hendrickson, Louise A. Brinton, Julie E. Buring, Hannia Campos, Qi Dai, Joanne F. Dorgan, Adrian A. Franke, Yu-tang Gao, Marc T. Goodman, Göran Hallmans, Kathy J. Helzlsouer, Judy Hoffman-Bolton, Kerstin Hultén, Howard D. Sesso, Anne L. Sowell, Rulla M. Tamimi, Paolo Toniolo, Lynne R. Wilkens, Anna Winkvist, Anne Zeleniuch-Jacquotte, Wei Zheng, Susan E. Hankinson

Manuscript received February 10, 2012; revised October 2, 2012; accepted October 4, 2012.

Correspondence to: A. Heather Eliassen, ScD, Channing Division of Network Medicine, Department of Medicine, Brigham & Women's Hospital and Harvard Medical School, 181 Longwood Ave, Boston, MA 02115 (e-mail: heather.eliassen@channing.harvard.edu).

- **Background** Carotenoids, micronutrients in fruits and vegetables, may reduce breast cancer risk. Most, but not all, past studies of circulating carotenoids and breast cancer have found an inverse association with at least one carotenoid, although the specific carotenoid has varied across studies.
 - **Methods** We conducted a pooled analysis of eight cohort studies comprising more than 80% of the world's published prospective data on plasma or serum carotenoids and breast cancer, including 3055 case subjects and 3956 matched control subjects. To account for laboratory differences and examine population differences across studies, we recalibrated participant carotenoid levels to a common standard by reassaying 20 plasma or serum samples from each cohort together at the same laboratory. Using conditional logistic regression, adjusting for several breast cancer risk factors, we calculated relative risks (RRs) and 95% confidence intervals (CIs) using quintiles defined among the control subjects from all studies. All *P* values are two-sided.
 - **Results** Statistically significant inverse associations with breast cancer were observed for α -carotene (top vs bottom quintile RR = 0.87, 95% Cl = 0.71 to 1.05, $P_{trend} = .04$), β -carotene (RR = 0.83, 95% Cl = 0.70 to 0.98, $P_{trend} = .02$), lutein+zeaxanthin (RR = 0.84, 95% Cl = 0.70 to 1.01, $P_{trend} = .05$), lycopene (RR = 0.78, 95% Cl = 0.62 to 0.99, $P_{trend} = .02$), and total carotenoids (RR = 0.81, 95% Cl = 0.68 to 0.96, $P_{trend} = .01$). β -Cryptoxanthin was not statistically significantly associated with risk. Tests for heterogeneity across studies were not statistically significant. For several carotenoids, associations appeared stronger for estrogen receptor negative (ER⁻) than for ER⁺ tumors (eg, β -carotene: ER⁻: top vs bottom quintile RR = 0.52, 95% Cl = 0.36 to 0.77, $P_{trend} = .001$; ER⁺: RR = 0.83, 95% Cl = 0.66 to 1.04, $P_{trend} = .06$; $P_{heterogeneity} = .01$).

J Natl Cancer Inst 2012;104:1905-1916

Carotenoids, natural pigments ranging from yellow to red, are important for photosynthesis in plants. More than 600 carotenoids have been identified (1), approximately 40 are present in the US diet, 20 of which are measurable in tissue and serum. α -Carotene, β -carotene, β -cryptoxanthin, lutein, zeaxanthin, and lycopene are the most prevalent, comprising 90% of circulating carotenoids (1–3). These micronutrients are suspected to be anticarcinogenic, with possible biologic activities including antioxidation, enhanced gap-junction intercellular communication, immunoenhancement, inhibition of tumorigenesis and malignant transformation, and metabolism to retinoids, which, in turn, contribute to cellular differentiation (4–10). Experimental studies suggest carotenoids inhibit tumor progression and reduce proliferation in both estrogen receptor positive (ER⁺) and ER⁻ breast cancer cells (11). Studies of dietary intake, of fruits and vegetables overall and carotenoids specifically, have had mixed results, although a modest inverse association has been suggested in several studies, as summarized in meta-analyses and pooled analyses (12–14). Several (15–19), but not all (20), studies of dietary patterns that include higher fruit and vegetable intake observed lower risk of ER⁻, but not ER⁺, breast tumors. Most recently, in pooled analyses of 18 cohort studies, stronger inverse associations were observed for ER⁻ (vs ER⁺) tumors with intake of fruits and vegetables (S Y Jung, D Spiegelman, L Baglietto, et al., unpublished data) as well as intake of α -carotene, β -carotene, and lutein+zeaxanthin (21).

Examining circulating carotenoid levels overcomes potential weaknesses of dietary data, including recall of past diet (3), inaccuracies of nutrient databases to determine carotenoid content of specific foods (22), influences of cooking and storage on carotenoid content (3,23), geographic and seasonal variation of foods (22), and individual differences in nutrient absorption. As a result of these limitations, the correlations between intake and plasma carotenoid levels are modest (r = 0.2-0.4) (24,25). Over the last 13 years, 10 prospective studies have investigated the associations between circulating carotenoids and breast cancer risk (26–35), eight of which are included in this analysis (26–33). Most studies observed inverse associations, either suggested or statistically significant, with at least one of five carotenoids (α -carotene, β -carotene, β -cryptoxanthin, lutein+zeaxanthin, lycopene), although the specific carotenoid and/ or subgroup of importance varied among studies.

The collaborating investigators of eight studies conducted a pooled analysis of original data examining associations between circulating carotenoids and breast cancer risk, the results of which are presented here.

Methods

A review of the literature at the time this project was proposed revealed five small, early studies (14 to 67 case subjects) (36–40) and six more recent, larger prospective studies (26–31). Investigators of the six larger studies were contacted, and all agreed to collaborate. During the data collection phase, an additional two studies were published and subsequently included in the collaborative project (32,33). The eight prospective studies in this pooled analysis are: Columbia, Missouri (26); Umea, Sweden (27); New York University Women's Health Study (NYUWHS), New York, New York (28); CLUE I and CLUE II, Washington County, Maryland (29); Nurses' Health Study (NHS), United States (30); Women's Health Study (WHS), United States (31); Shanghai Women's Health Study (SWHS), Shanghai, China (32); and Multiethnic Cohort Study (MEC), California and Hawaii (33). Two studies were published after the analyses were underway, and were not included in this pooled analysis (34,35).

Each of the eight studies was a case–control study nested within a cohort, with plasma (27,29–33) or serum (26,28,29) samples collected among initially healthy women who were then followed for subsequent risk of breast cancer. Investigators of the original studies contributed data on circulating carotenoid concentrations, matching factors, covariates, and breast cancer diagnoses. Each study obtained informed consent (or proxy, such as implied consent with return of blood samples and questionnaires in the NHS) and approval from their respective institutional review boards. The pooled analysis was approved by the Committee on the Use of Human Subjects in Research at the Brigham and Women's Hospital.

Carotenoid Assays

Reverse-phase high-performance liquid chromatography was used to measure plasma or serum carotenoids in each of the original studies, but assays were conducted at five different laboratories. To distinguish between-laboratory variation from between-population variation, we conducted a recalibration study. Plasma (27,29–33) or serum (26,28,29) samples from each study that had been assayed for carotenoids in the original analysis were sent to the NHS blood laboratory. Samples were from control subjects for seven of the eight studies; quality control samples were used for the remaining study (MEC). Control subject samples were chosen to

represent a range of carotenoid levels, with selection of two control subjects from each study-specific quintile of α -carotene and total carotenoids. The original data in the NHS showed considerable laboratory drift across four batches run in different years (30); thus, 20 control subjects from each of four batches were sent for recalibration. Samples from seven of eight studies (not SWHS) were realiquoted, labeled to blind the assay laboratory to the study identification, and packaged together in one project with additional quality control samples and 10 "drift pool" samples, which are additional unique quality control samples used to assess drift among batches. SWHS provided samples (n = 10) that were assayed in a separate, later batch but with identical quality control and drift pool samples as were sent with the other study samples. Samples were assayed by reverse-phase high-performance liquid chromatography, using the methods described by El-Sohemy et al. (41), at the Micronutrient Analysis Laboratory in the Department of Nutrition at the Harvard School of Public Health. Individual carotenoid measures include all specific isomers (eg, cis+trans); lutein and zeaxanthin are read together, and therefore are a sum of the two individual carotenoids (lutein+zeaxanthin). Coefficients of variation from blinded quality control replicates ranged from 4.6% (lutein+zeaxanthin) to 8.2% (α -carotene).

Matching Factor and Covariate Data

Information was collected on matching factors, measured plasma or serum carotenoid levels, and case-control status. Matching factors varied across cohorts, with cohorts matching on three to seven factors, including age at blood collection; date, time, and fasting status at blood collection; menopausal status; date of last menstrual period and/or phase and day of menstrual cycle in premenopausal women; postmenopausal hormone use; race or ethnicity; study center; smoking status; follow-up time; availability of food frequency questionnaire; use of antibiotics in the last week; number of blood collections within the cohort; and diagnosis of benign breast disease in the previous 2 years. Matches on date of blood collection included ± 1 month (CLUE I, CLUE II, NHS, SWHS), ± 2-3 months (Umea, Sweden; New York University Women's Health Study), ±6 months (MEC), and ± 1 year (Columbia, MO). Although blood collection date was not a matching factor in WHS, case subjects were matched to control subjects on follow-up time (±6 months), and 80% of case-control pairs had blood collection dates within 10 months (all were within 2 years).

To comprehensively adjust for possible confounders and allow for stratified analyses, we requested information from each study for the case and control subjects on age at menarche, parity, age at first birth, menopausal status, age at menopause, body mass index (BMI), family history of breast cancer, personal history of benign breast disease, smoking status, alcohol use, oral contraceptive use, postmenopausal hormone use, cholesterol level, physical activity, and race/ethnicity. The NHS and WHS cohorts had information available on each of these covariates. Data from each cohort was available for all variables except BMI, personal history of benign breast disease, alcohol use, cholesterol level, and physical activity. A range of one (CLUE II, SWHS) to four (CLUE I) variables was missing from each of the remaining cohorts.

Covariates were coded with as much detail as possible within the restriction of creating common definitions across cohorts.

Downloaded from https://academic.oup.com/jnci/article-abstract/104/24/1905/920659 by guest on 31 May 2018 Multiple cohorts were missing data on physical activity (5 cohorts), alcohol consumption (4 cohorts), and cholesterol levels (2 cohorts).

Case Subjects

From each cohort, we attempted to collect diagnosis information, including date, hormone receptor status, differentiation, size, and nodal involvement. A total of 3055 case subjects were included, matched with 3956 control subjects. Seven cohorts had information on hormone receptor status ($n = 1481 \text{ ER}^+$ case subjects), $n = 417 \text{ ER}^-$ case subjects). We obtained information on grade from six cohorts (n = 1479 case subjects), information of size from four cohorts (n = 1617 case subjects), and information on nodal involvement from six cohorts (n = 1810 case subjects).

Statistical Analysis

Outliers, based on the original carotenoid values, were detected within each cohort using the extreme Studentized many-deviate approach (42). Outliers were removed for the individual carotenoids, with the sum of outliers across cohorts ranging from 2 to 17 for each carotenoid.

Original circulating carotenoid values were recalibrated to have a distribution comparable with the samples rerun together at the Harvard laboratory. Using linear regression within each cohort, we regressed the rerun values on the original values and used the resulting intercept and slope to predict recalibrated values for all cohort participants. Quintiles and deciles of individual and total carotenoids were defined among control subjects, within and across studies, using both original and recalibrated values, thus resulting in cut points classified in four ways: study-specific original values, study-specific recalibrated values.

Analyses were conducted in two ways, by using a two-stage approach of pooled relative risks (RRs) and by an aggregated data approach of pooled raw data. In the two-stage approach, naturallog relative risks were calculated from conditional logistic regression models within each cohort and pooled using DerSimonian and Laird random effects models (43), with relative risks weighted by the inverse of their variance. We tested for heterogeneity among studies by the DerSimonian and Laird *Q* statistic (43). In the aggregated data approach, we pooled the raw data from each cohort and calculated a single, combined relative risk from a conditional logistic regression model (44).

Multivariable adjustment was achieved by considering the most thorough model with the least amount of missing information. Our final, multivariable-adjusted, conditional logistic regression model included the following variables: menopausal status (premenopausal, postmenopausal, unknown); age at menopause (years, continuous); age at menarche (≤ 12 , 13, ≥ 14 years, missing); parous (yes, no); age at first birth (years, continuous); exogenous hormone use (oral contraceptives or postmenopausal hormones; yes, no, missing); BMI (kg/m², continuous); current smoking (yes, no, missing); race (Caucasian, African-American, Asian/Pacific Islander, other, missing); personal history of benign breast disease (yes, no, missing); and family history of breast cancer (yes, no, missing). Although some of these variables were matching factors in some studies (eg, menopausal status was a matching factor in five studies), we included them as covariates to ensure we achieved control for possible confounding in the combined analysis. To assess the effect of simplifying variable definitions, we conducted sensitivity analyses within NHS and WHS, cohorts with the most detailed covariate information, comparing detailed vs simplified coding in the multivariable-adjusted models; results were not appreciably different. In addition, we conducted sensitivity analyses including vs excluding physical activity, alcohol consumption, and cholesterol levels within the cohorts that had information on these variables; results generally were similar.

For stratified analyses, we used unconditional logistic regression, additionally adjusting for the following matching factors: age (years), date of blood collection (months), time of blood collection (hours, missing), fasting status (yes, no, missing), and study [12 indicators including CLUE I and II separately as well as analytic subsets within Umea, Sweden (27) and MEC (33)]. We conducted analyses stratified by lifestyle factors (menopausal status [premenopausal vs postmenopausal], postmenopausal hormone use [current use, yes vs no], smoking status [current, yes vs no], alcohol consumption [<4 vs \geq 4 drinks per week], BMI [<25, 25 to <30, \geq 30 kg/m²], and time between blood collection and diagnosis [<2, 2–5, \geq 5 years]) and by tumor characteristics (size [<2 vs \geq 2 cm], nodal involvement [0 vs \geq 1 positive nodes], differentiation [well, moderate, poor], and ER status [ER⁺ vs ER⁻]).

Tests for trend were conducted by modeling the medians of quintiles (or deciles) as a continuous variable and calculating the Wald statistic. The shape of the dose-response curves and tests for nonlinearity were assessed using restricted cubic spline models with stepwise selection of knots over the range of carotenoid distributions (45). Wald tests for interaction between stratification variables and carotenoids compared the slopes of the quintile medians between groups. To test whether associations differed by tumor characteristics, we used polychotomous logistic regression with multiple endpoints (eg, ER+, ER-, and no breast cancer) (46). We used a likelihood ratio test to compare a model with separate slopes for the carotenoids in each case group with a model with a common slope. Correlations with plasma or serum carotenoid levels were assessed by Spearman rank correlations. All P values were based on two-sided tests and were considered statistically significant if less than or equal to .05. Analyses were conducted using SAS version 9 (SAS Institute, Cary, NC); polychotomous logistic regressions were conducted using STATA version 11.0 (StataCorp, College Station, TX).

Results

Each of the eight studies included in the pooled analysis contributed a range of 105 to 962 breast cancer case subjects (total = 3055) and a range of 115 to 962 control subjects (total = 3956) (Table 1). Mean age at blood collection for case subjects ranged across studies from 51.3 to 66.0 years. Most participants were postmenopausal at blood collection within studies (50%-100%) and overall (67%). Median time between blood collection and diagnosis was 4.3 years (range = 0.8-13.7 years).

Recalibration of original plasma or serum carotenoid values resulted in consistent, high correlations between the original and recalibrated carotenoids values. The regression models of original and rerun carotenoid levels for each carotenoid resulted in

		e	
Table 1. Case and control characteristics in	nooled analysis o	t prospective data on circulati	nd carotenoids and breast cancer risk*
	poolea analysis o		ing carotonolas ana breast cancer hisk

Cohort	Reference	Subjects	No.	Mean age, y	Mean BMI, kg/m²	Mean age menarche, y	Nulli- parous, %	Postmeno- pausal, %	0	Median time to diagnosis, y
Columbia, MO	Dorgan et al., 1998 (26)	Case	105	58.2	26.1	12.9	18.1	77.1	23.5	2.7
		Control	209	58.2	26.8	12.9	14.4	79.4	20.5	_
Umea, Sweden Hultén et al., 200	Hultén et al., 2001 (27)	Case	201	54.5	25.3	13.3	12.7	66.7	45.5	0.8
		Control	389	54.4	25.7	13.4	9.8	66.6	40.2	_
CLUE I Sato et al., 2002 (2	Sato et al., 2002 (29)	Case	244	51.3	‡	12.7	1.1	57.0	13.0	13.7
		Control	244	51.1	‡	12.7	1.3	57.8	14.2	_
CLUE II Sato	Sato et al., 2002 (29)	Case	115	60.4	26.2	12.6	1.1	79.1	11.0	2.4
		Control	115	60.2	25.4	12.9	1.1	79.1	13.2	_
NHS Tamimi et al., 20	Tamimi et al., 2005 (30)	Case	962	57.2	25.4	12.5	6.4	68.6	52.6	4.5
		Control	962	57.3	25.4	12.6	6.6	68.3	52.7	_
WHS Se	Sesso et al., 2005 (31)	Case	508	55.7	25.5	12.3	2.3	62.4	59.3	3.7
		Control	508	55.7	25.9	12.4	2.6	61.0	54.2	_
NYUWHS	Toniolo et al., 2006 (28)	Case	269	53.4	25.3	12.4	34.6	53.2	0	3.7
		Control	269	53.2	25.4	12.6	29.4	53.2	0	_
SWHS Dorjgochoo et al.,	Dorjgochoo et al., 2009 (32)	Case	365	53.1	24.3	14.8	4.1	49.6	7.7	3.5
		Control	725	53.2	24.5	15.0	3.5	50.6	3.3	_
MEC E	Epplein et al., 2009 (33)	Case	286	66.0	25.6	12.5	12.2	100.0	34.6	1.5
		Control	535	66.0	25.4	12.6	10.8	100.0	34.6	_
Total		Case	3055	56.5	25.3	12.8	9.0	66.5	37.2	4.3
		Control	3956	56.7	25.4	13.1	8.1	67.5	33.0	

* BMI = body mass index; MEC = Multiethnic Cohort Study; NHS = Nurses' Health Study; NYUWHS = New York University Women's Health Study; PMH = postmenopausal hormones; SWHS = Shanghai Women's Health Study; WHS = Women's Health Study; — = not applicable because controls were not diagnosed with breast cancer.

† Using PMH calculated among postmenopausal women only.

‡ BMI not collected in CLUE I.

relatively high R^2 values from individual studies (α -carotene = 0.89-0.99; β -carotene = 0.86-0.99; β -cryptoxanthin = 0.92-0.99; lutein+zeaxanthin = 0.74-0.98; lycopene = 0.80-0.99) with the exception of WHS ($R^2 = 0.56$) and MEC ($R^2 = 0.18$). There was no obvious explanation for the low lycopene correlations in WHS and MEC. Although three high original values in the WHS appeared to influence the regression, and excluding them increased the R^2 to 0.85, recalibration results were similar whether including or excluding these values. Although correlations generally were high, absolute values of original and recalibrated individual carotenoid values differed with recalibration (Figure 1; Supplementary Table 1, available online). In general, differences among studies were not as pronounced with recalibration. For example, the range of median values of α -carotene narrowed from the original (range = $1.7-12.7 \ \mu g/dL$) to the recalibrated ($2.3-8.9 \ \mu g/dL$) values. In most instances, the ranking of medians across studies was maintained with recalibration. The exceptions include β-carotene values in three studies (Umea, Sweden, from middle to high; Columbia, MO, from high to lowest; MEC, from highest to middle with recalibration), β-cryptoxanthin and lutein+zeaxanthin in NHS (from low to middle-high with recalibration), and lycopene in WHS (from second lowest to second highest with recalibration). Recalibrated values highlight population differences in carotenoid levels between studies. For example, the 90th percentile of α -carotene in SWHS is nearly comparable with the 10th percentile in WHS; the 90th percentiles of β -cryptoxanthin in CLUE I and CLUE II are comparable with the median in MEC (Supplementary Table 1, available online). In SWHS, the 10th percentile of lutein+zeaxanthin is higher than the medians of all

but two cohorts (NHS and New York University Women's Health Study); in contrast, the median of lycopene in SWHS is lower than the 10th percentiles of all but two cohorts (Columbia, MO, and Umea, Sweden).

We first examined associations between carotenoid levels and breast cancer risk using the two-stage approach, with original carotenoid values and study-specific quintiles (data not shown). Suggestive but non-statistically significant inverse associations were observed for α -carotene (top vs bottom quintile RR= 0.84, 95% CI = 0.63 to 1.10, P_{trend} = .19); β -carotene (RR = 0.77, 95%) CI = 0.56 to 1.05, P_{trend} = .16), and total carotenoids (RR = 0.72, 95% CI = 0.51 to 1.01, P_{trend} = .07). However, statistically significant heterogeneity between studies was observed for each of these carotenoids (P = .03,.004,.002, respectively). β -Cryptoxanthin was not associated with breast cancer risk (RR = 0.98, 95% CI = 0.83 to 1.17, $P_{\text{trend}} = .67$). Associations for lutein+zeaxanthin (RR = 0.91, 95% CI = 0.77 to 1.07, P_{trend} = .24) and lycopene (RR = 0.89, 95% CI = 0.72 to 1.08, $P_{trend} = .01$) were similar to one another. Tests for heterogeneity were not statistically significant for β -cryptoxanthin, lutein+zeaxanthin, or lycopene.

Using the recalibrated data and common quintiles across cohorts, we conducted analyses with the aggregated data approach (Figure 2). In unadjusted conditional logistic regression models, we observed suggestive inverse associations with some carotenoids and a statistically significant inverse trend for lycopene (top vs bottom quintile RR = 0.80, 95% CI = 0.64 to 1.01, P_{trend} = .05). Associations strengthened with multivariable adjustment because of small contributions by several factors (eg, BMI, parity, smoking, menopausal status, and postmenopausal hormone use). Statistically significant inverse

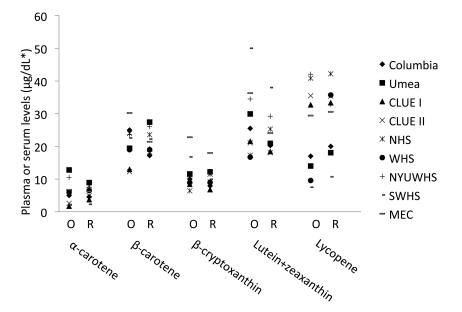


Figure 1. Original (O) and recalibrated (R) medians of plasma or serum levels of carotenoids by study, among control subjects. *To convert μ g/dL to μ mol/L, multiply by the following factors: 0.01863 for α -carotene, β -carotene, and lycopene; 0.01810 for β -cryptoxanthin; 0.01758 for lutein+zeaxanthin. MEC = Multiethnic Cohort Study; NHS = Nurses' Health Study; NYUWHS = New York University Women's Health Study; SWHS = Shanghai Women's Health Study; WHS = Women's Health Study.

associations were observed for all carotenoids except ß-cryptoxanthin (α -carotene: top vs bottom quintile RR = 0.87, 95% CI = 0.71 to 1.05; $P_{\text{trend}} = .04$; β -carotene: RR = 0.83, 95% CI = 0.70 to 0.98, P_{trend} = .02; β -cryptoxanthin: RR = 0.98, 95% CI = 0.82 to 1.18, P_{trend} = .21; lutein+zeaxanthin: RR = 0.84, 95% CI = 0.70 to 1.01, $P_{\text{trend}} = .05$; lycopene: RR = 0.78, 95% CI = 0.62 to 0.99, P_{trend} = .02; total carotenoids: RR = 0.81, 95% CI = 0.68 to 0.96, P_{trend} = .01). None of the associations was statistically significantly nonlinear in cubic spline models (data not shown). We reran the model assessing the association with lycopene excluding MEC and/or WHS given the low correlations between original and rerun assay levels; results essentially were unchanged (data not shown). Using the two-stage approach with recalibrated data and common quintiles, results generally were similar to the aggregated data approach (data not shown). Tests for heterogeneity among cohorts were no longer statistically significant (data not shown). All subsequent analyses were conducted using the aggregated data approach with recalibrated carotenoid levels and common cut points.

Expanding the quantile analysis to deciles, we generally observed that breast cancer risk continued to decrease in the top decile of carotenoids (α -carotene: top vs. bottom decile RR = 0.79, 95% CI = 0.61 to 1.03, $P_{\text{trend}} = .02$; β -carotene: RR = 0.77, 95% CI = 0.61 to 0.98, $P_{\text{trend}} = .01$; lutein+zeaxanthin: RR = 0.72, 95% CI = 0.56 to 0.92, $P_{\text{trend}} = .02$; total carotenoids: RR = 0.71, 95% CI = 0.56 t 0.91, $P_{\text{trend}} = .003$). The association between lycopene and breast cancer risk was not as consistent across deciles as it was across quintiles (top vs bottom decile RR = 0.89, 95% CI = 0.65 to 1.21, $P_{\text{trend}} = .11$).

When we analyzed the associations between carotenoids and breast cancer risk by ER status, inverse associations were apparent among ER⁻ tumors (Figure 3). Although inverse associations were suggested among ER⁺ tumors for α -carotene and β -carotene, multivariable relative risks and tests for trend were not statistically significant (top vs bottom quintile RR = 0.85, 95% CI = 0.65 to 1.12, $P_{\text{trend}} = .16$; RR = 0.83, 95% CI = 0.66 to 1.04; $P_{\text{trend}} = .06$, respectively). However, the associations were statistically significant for both α -carotene and β -carotene among ER⁻ tumors (top vs bottom quintile RR = 0.61, 95% CI = 0.40 to 0.93, $P_{\text{trend}} = .04$; RR = 0.52, 95% CI = 0.36 to 0.77, $P_{\text{trend}} = .001$, respectively). There was statistically significant heterogeneity between ER⁻ and ER⁺ tumors for β -carotene (P = .01) but not for α -carotene (P = .11). Associations for other carotenoids were not statistically significantly different by ER status.

We compared associations between carotenoids and breast cancer risk across categories of tumor size, nodal involvement, and differentiation. No statistically significant differences were observed for tumor size (<2 vs ≥2cm) or nodal involvement (0 vs ≥1 positive nodes) (data not shown). A statistically significant difference by differentiation was observed for lutein+zeaxanthin, with stronger inverse associations with less differentiation (well-differentiated (n = 301): top vs bottom quintile RR = 1.35, 95% CI = 0.87 to 2.09, P_{trend} = .28; moderately differentiated (n = 664): RR = 0.88, 94% CI = 0.64 to 1.23, P_{trend} = .28; poorly differentiated (n = 508): RR = 0.68, 95% CI = 0.46 to 0.99, P_{trend} = .15; $P_{\text{heterogeneity}}$ = .04). Differences in the associations by differentiation were not observed for any other carotenoid.

We conducted analyses stratified by several lifestyle factors, including menopausal status, postmenopausal hormone use, smoking status, alcohol consumption, BMI, and time between blood collection and diagnosis. Interactions with most of these factors were not statistically significant. However, several of the associations of carotenoid with breast cancer risk varied by BMI and smoking status (Table 2). Interactions with BMI were fairly consistent across carotenoids, with stronger inverse associations with breast cancer risk observed among leaner (BMI ≤25) women and suggestively or statistically significantly positive associations among obese women

Downloaded from https://academic.oup.com/jnci/article-abstract/104/24/1905/920659 by guest on 31 May 2018

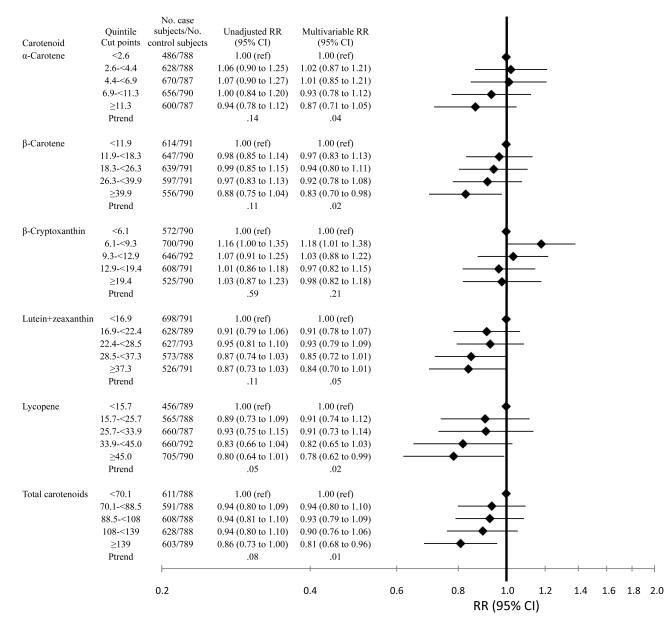


Figure 2. Relative risks (RRs) of breast cancer and 95% confidence intervals (CIs) according to quintile of plasma carotenoids (μ g/dL), recalibrated data. **Diamonds** represent relative risks; **lines** represent 95% confidence intervals. To convert μ g/dL to μ mol/L, multiply by the following factors: 0.01863 for α -carotene, β -carotene, and lycopene; 0.01810 for β -cryptoxanthin; 0.01758 for lutein+zeaxanthin. ref = referent.

(BMI = ≥30) (eg, total carotenoids: BMI <25: top vs bottom quartile RR = 0.65, 95% CI = 0.51 to 0.83, P_{trend} = .001; BMI ≥30: RR = 1.74, 95% CI = 0.97 to 3.13, P_{trend} = .04; $P_{\text{heterogeneity}}$ = .01). Interestingly, as has been documented previously (3), BMI was inversely associated with plasma carotenoid levels (total carotenoids r = -0.19).

Statistically significant interactions also were observed with current smoking for lutein+zeaxanthin and total carotenoids, with stronger inverse associations among current smokers (Table 2) (eg, total carotenoids: non-smokers: top vs bottom quintile RR = 0.93, 95% CI = 0.77 to 1.11, $P_{\text{trend}} = .38$; current smokers: RR = 0.47, 95% CI = 0.30 to 0.73, $P_{\text{trend}} = .002$; $P_{\text{heterogeneity}} = .01$). Although interactions with smoking were not statistically significant for the remaining carotenoids, generally similar patterns were observed,

with stronger associations among smokers. This pattern persisted among both lean and heavy women (data not shown). Similarly, the interaction with BMI persisted among nonsmokers (data not shown).

Statistically significant interactions with smoking persisted regardless of ER status for lutein+zeaxanthin and total carotenoids (data not shown). Although interactions with BMI were not statistically significant, differences in the carotenoids associations by BMI appeared more striking among ER⁻ tumors than among ER⁺ tumors (eg, among ER⁻ tumors: α -carotene BMI <25: top vs bottom quintile RR = 0.34, 95% CI = 0.18 to 0.64; vs BMI ≥25: RR = 1.06, 95% CI = 0.54 to 2.09; among ER⁺ tumors: α -carotene BMI <25: RR = 0.80, 95% CI =0.51 to 1.24; vs BMI >25: RR = 1.13, 95% CI = 0.75 to 1.70). Although we observed

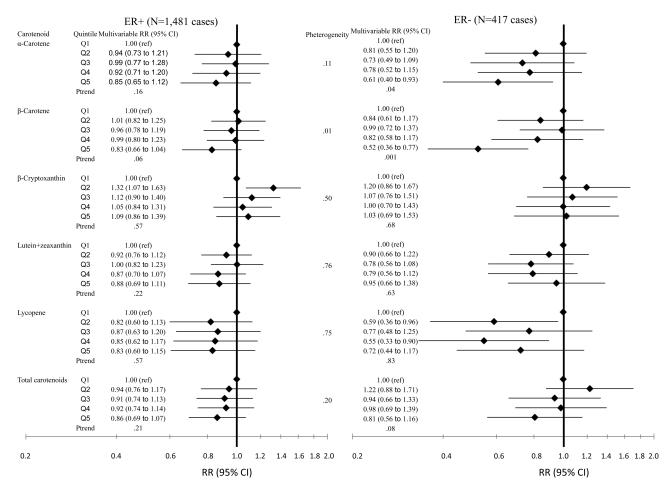


Figure 3. Relative risks (RRs) of breast cancer and 95% confidence intervals (CIs) according to quintile of plasma carotenoids, recalibrated data, by tumor estrogen receptor (ER) status. **Diamonds** represent relative risks; **lines** represent 95% confidence intervals. ER⁺ = estrogen receptor positive; ER⁻ = estrogen receptor negative; ref = referent.

no association between β -cryptoxanthin and breast cancer risk overall and by ER status, we observed strong inverse associations with ER⁻ tumors among lean women and strong positive associations with ER⁻ tumors among overweight and obese women (BMI <25: top vs. bottom quintile RR = 0.50, 95% CI = 0.28 to 0.88, $P_{\text{trend}} = .03$; BMI ≥25: RR =2.18, 95% CI = 1.13 to 4.24, $P_{\text{trend}} = .07$; $P_{\text{heterogeneity}} = .16$).

To investigate the role of vitamin supplement use in the observed associations, we repeated the analyses among six cohorts with data on multivitamin and/or supplement use and restricted analyses to nonusers in these cohorts. Overall results among these cohorts (n = 2749 case subjects) were similar to the total pooled results. Restricting to nonvitamin users (n = 1364 case subjects) generally resulted in similar point estimates but non-statistically significant trends, except for lutein+zeaxanthin and lycopene, for which the point estimates were somewhat attenuated as well as non-statistically significant (data not shown).

To examine the potential impact of preclinical disease on our results, we excluded 897 case subjects diagnosed less than or equal to 2 years after blood collection, ranging from 4% (CLUE I) to 69% (Umea, Sweden) of case subjects within individual cohorts. Risk estimates were essentially unchanged for α -carotene, β -carotene, β -cryptoxanthin, and total carotenoids, but somewhat attenuated for lutein+zeaxanthin and lycopene (data not shown).

The levels of most individual carotenoids were statistically significantly correlated with one another, ranging from r equal to 0.14 (P < .001) (lutein+zeaxanthin with α -carotene) to r equal to 0.65 (P < .001) (α -carotene with β -carotene); lutein+zeaxanthin and lycopene were not correlated (r = -0.02, P = .27). When pairs of carotenoids were included in the same statistical model, most associations were attenuated and no longer statistically significant (eg, β -carotene adjusting for α -carotene: top vs bottom quintile RR = 0.87, 95% CI = 0.70 to 1.08, P_{trend} = .19) except when β -cryptoxanthin was the second carotenoid (eg, β -carotene adjusting for β -cryptoxanthin: RR = 0.85, 95% CI = 0.71 to 1.02, $P_{\text{trend}} = .05$). The inverse association between lycopene and breast cancer risk remained statistically significant with adjustment for either β -cryptoxanthin or lutein+zeaxanthin (eg, lycopene: RR = 0.80, 95% CI = 0.63 to 1.02, P_{trend} = .05 adjusting for lutein+zeaxanthin). Point estimates for the association between lycopene and breast cancer were similar to the overall point estimates when adjusting for α -carotene (correlation with lycopene: r = 0.42, P < .001) or β -carotene (r = 0.25, P < .001), but the trends were no longer statistically significant (eg, RR = 0.81, 95% CI = 0.64 to 1.03, $P_{\text{trend}} = .06$ adjusting for α -carotene).

Downloaded from https://academic.oup.com/jnci/article-abstract/104/24/1905/920659 by guest on 31 May 2018

Table 2. Multivariable relative risk (95% confidence interval) of breast cancer according to quintile of circulating carotenoids, recalibrated
data, overall and stratified by body mass index (BMI) or smoking*

Carotenoid	Case subjects	Control subjects	Q1	02	Q3	Q4	Ω5	P _{trend}	$P_{\rm het}$
α-Carotene									
Overall	3040	3940	1.00 (ref)	1.02 (0.87 to 1.21)	1.01 (0.85 to 1.21)	0.93 (0.78 to 1.12)	0.87 (0.71 to 1.05)	.04	
BMI <25	1522	1978	1.00 (ref)	1.01 (0.78 to 1.31)	1.04 (0.79 to 1.36)	0.96 (0.73 to 1.26)	0.77 (0.58 to 1.02)	.004	
BMI 25 to <30	875	1188	1.00 (ref)	1.23 (0.89 to 1.69)	1.11 (0.80 to 1.54)	0.91 (0.65 to 1.28)	1.11 (0.78 to 1.57)	.83	
BMI ≥30	387	487	1.00 (ref)	0.77 (0.50 to 1.17)	1.10 (0.71 to 1.71)	1.45 (0.89 to 2.37)	1.36 (0.76 to 2.45)	.05	.01
Nonsmokers	2531	3317	1.00 (ref)	1.09 (0.91 to 1.32)	1.08 (0.89 to 1.32)	1.04 (0.84 to 1.27)	0.94 (0.76 to 1.16)	.13	
Current smokers	485	592	1.00 (ref)	0.85 (0.58 to 1.25)	0.84 (0.56 to 1.26)	0.87 (0.56 to 1.35)	0.76 (0.48 to 1.23)	.39	.90
β-Carotene									
Overall	3053	3953	1.00 (ref)	0.97 (0.83 to 1.13)	0.94 (0.80 to 1.11)	0.92 (0.78 to 1.08)	0.83 (0.70 to 0.98)	.02	
BMI <25	1523	1978	1.00 (ref)	0.95 (0.74 to 1.23)	0.86 (0.67 to 1.11)	0.89 (0.70 to 1.14)	0.72 (0.56 to 0.91)	.002	
BMI 25 to <30	875	1190	1.00 (ref)	1.07 (0.81 to 1.40)	0.96 (0.73 to 1.27)	1.03 (0.77 to 1.38)	1.10 (0.81 to 1.50)	.58	
BMI ≥30	388	488	1.00 (ref)	0.81 (0.56 to 1.19)	1.65 (1.09 to 2.49)	0.93 (0.58 to 1.50)	1.35 (0.73 to 2.51)	.27	.01
Nonsmokers	2537	3325	1.00 (ref)	1.03 (0.87 to 1.23)	1.06 (0.89 to 1.26)	1.02 (0.86 to 1.22)	0.91 (0.76 to 1.09)	.16	
Current smokers	492	597	1.00 (ref)	0.75 (0.52 to 1.07)		0.72 (0.47 to 1.08)	0.57 (0.36 to 0.90)	.03	.07
β-Cryptoxanthin	102	007		01/0 (0102 10 110/)	0.00 (0.11 10 0.00)				,
Overall	3051	3953	1.00 (ref)	1.18 (1.01 to 1.38)	1.03 (0.88 to 1.22)	0.97 (0.82 to 1.15)	0.98 (0.82 to 1.18)	.21	
BMI <25	1523	1977	1.00 (ref)	1.14 (0.89 to 1.46)	0.99 (0.77 to 1.27)	0.93 (0.72 to 1.19)	0.92 (0.72 to 1.18)	.15	
BMI 25 to <30	873	1191	1.00 (ref)	1.21 (0.91 to 1.62)	1.14 (0.85 to 1.54)	0.99 (0.73 to 1.35)	1.11 (0.80 to 1.55)	.92	
BMI ≥30	386	488	1.00 (ref)	1.19 (0.82 to 1.74)	1.14 (0.75 to 1.74)	0.87 (0.52 to 1.44)	1.59 (0.91 to 2.76)	.26	.05
Nonsmokers	2536	3325	1.00 (ref)	1.13 (0.95 to 1.34)	1.02 (0.86 to 1.22)	0.99 (0.83 to 1.19)	1.02 (0.85 to 1.23)	.63	.00
Current smokers	491	597	1.00 (ref)	1.27 (0.90 to 1.80)	1.12 (0.77 to 1.64)	0.95 (0.62 to 1.46)	0.68 (0.39 to 1.19)	.00	.21
Lutein+zeaxanthin	401	007	1.00 (101)	1.27 (0.00 to 1.00)	1.12 (0.77 to 1.04)	0.00 (0.02 to 1.40)	0.00 (0.00 to 1.10)	.00	.21
Overall	3052	3952	1.00 (ref)	0.91 (0.78 to 1.07)	0.93 (0.79 to 1.09)	0.85 (0.72 to 1.01)	0.84 (0.70 to 1.01)	.05	
BMI <25	1523	1977	1.00 (ref)	0.70 (0.55 to 0.90)			0.63 (0.49 to 0.82)	.004	
BMI 25 to <30	873	1191	1.00 (ref)	0.93 (0.70 to 1.24)	0.83 (0.62 to 1.10)	0.88 (0.65 to 1.19)	1.00 (0.72 to 1.39)	.99	
BMI ≥30	387	487	1.00 (ref)	1.42 (0.97 to 2.07)	1.22 (0.80 to 1.85)	1.02 (0.63 to 1.64)	1.89 (1.03 to 3.47)	.15	.02
Nonsmokers	2537	3323	1.00 (ref)	0.89 (0.75 to 1.05)	0.92 (0.77 to 1.10)	0.91 (0.76 to 1.09)	0.88 (0.73 to 1.07)	.34	.02
Current smokers	491	598	1.00 (ref)	0.98 (0.69 to 1.39)	1.00 (0.69 to 1.44)	0.58 (0.37 to 0.90)		.04	.04
Lycopene	401	000	1.00 (101)	0.00 (0.00 to 1.00)	1.00 (0.03 to 1.44)	0.00 (0.07 to 0.00)	0.00 (0.00 to 0.00)	.01	.04
Overall	3046	3946	1.00 (ref)	0.91 (0.74 to 1.12)	0.91 (0.73 to 1.14)	0.82 (0.65 to 1.03)	0.78 (0.62 to 0.99)	.02	
BMI <25	1518	1974	1.00 (ref)	0.90 (0.68 to 1.18)	1.03 (0.77 to 1.37)	0.91 (0.68 to 1.22)	0.81 (0.60 to 1.09)	.12	
BMI 25 to <30	873	1188	1.00 (ref)	0.84 (0.59 to 1.18)	0.79 (0.55 to 1.15)	0.75 (0.51 to 1.09)	0.76 (0.51 to 1.12)	.23	
BMI ≥30 BMI ≥30	386	487	1.00 (ref)	1.09 (0.63 to 1.87)	1.06 (0.59 to 1.89)	1.08 (0.60 to 1.96)	1.47 (0.80 to 2.72)	.23	.12
Nonsmokers	2530	3320	1.00 (ref)	0.93 (0.75 to 1.14)	1.00 (0.33 to 1.33) 1.01 (0.81 to 1.26)	0.89 (0.71 to 1.12)	0.90 (0.71 to 1.13)	.31	.12
Current smokers	492	595	1.00 (ref)	0.86 (0.54 to 1.37)		0.68 (0.41 to 1.12)	0.63 (0.38 to 1.05)	.06	.66
Total carotenoids	432	090	1.00 (101)	0.00 (0.04 (0 1.37)	0.05 (0.40 to 1.07)	0.00 (0.41 (0 1.12)	0.03 (0.38 t0 1.03)	.00	.00
Overall	3041	3941	1.00 (ref)	0.94 (0.80 to 1.10)	0.93 (0.79 to 1.09)	0.90 (0.76 to 1.06)	0.81 (0.68 to 0.96)	.01	
BMI <25	1523	1977	1.00 (ref)	0.80 (0.62 to 1.04)	0.82 (0.64 to 1.06)	0.80 (0.63 to 1.02)	0.65 (0.51 to 0.83)	.001	
BMI 25 to <30	874	1977	1.00 (ref) 1.00 (ref)	1.02 (0.77 to 1.36)	0.82 (0.64 to 1.06) 0.97 (0.73 to 1.28)	1.01 (0.76 to 1.36)	1.04 (0.76 to 1.42)	.001	
BMI 25 to <30 BMI ≥30	874 388	488							01
			1.00 (ref)	1.31 (0.90 to 1.90)	1.31 (0.85 to 2.00)	1.40 (0.88 to 2.24)	1.74 (0.97 to 3.13)	.04	.01
Nonsmokers	2532	3316	1.00 (ref)	1.00 (0.84 to 1.20)	0.95 (0.80 to 1.14)	0.98 (0.82 to 1.17)	0.93 (0.77 to 1.11)	.38	01
Current smokers	485	594	1.00 (ref)	0.74 (0.51 to 1.06)	1.00 (0.68 to 1.48)	0.78 (0.52 to 1.18)	0.47 (0.30 to 0.73)	.002	.01

* Models adjusted for menopausal status (premenopausal, postmenopausal, dubious status/unknown); age at menopause (years); age at menarche (≤12, 13, ≥14, missing); parous (yes, no); age at first birth (years); exogenous hormone use (oral contraceptives or postmenopausal hormones; yes, no, missing); BMI (kg/m²); race (Caucasian, African American, Asian/Pacific Islander, other, missing); personal history of benign breast disease (yes, no, missing); family history of breast cancer (yes, no, missing). For stratified analyses, we used unconditional logistic regression, additionally adjusting for the following matching factors: age (years), date of blood collection (months), time of blood collection (hours), fasting status (yes, no, missing), and study (12 indicators including subdivisions within Umea, Sweden, and Multiethnic Cohort Study); models stratified by BMI additionally were adjusted for current smoking (yes, no, missing). All *P* values are two-sided. *P*_{het} = *P* heterogeneity; **Q** = quintile; ref = referent.

Discussion

In this large pooled analysis with more than 3000 case subjects, we observed statistically significant inverse associations between circulating levels of individual and total carotenoids and breast cancer risk. Specifically, inverse associations were observed for α -carotene, β -carotene, lutein+zeaxanthin, and lycopene, but not β -cryptoxanthin. Associations generally were stronger among lean women and for ER⁻ tumors, and, for lutein+zeaxanthin and total carotenoids, associations were stronger among current smokers.

To date, four initial small (N = 14–67 case subjects) (36–40) and 10 larger, more recent (26–35) nested case–control studies of circulating carotenoid levels and breast cancer risk have been published. In most (26–30,32,35), but not all (31,33,34), of the larger studies, statistically significant or suggestive inverse associations were observed, with relative risk less than or equal to 0.7 for at least one carotenoid. Two studies (34,35) were published after this project was underway; thus, the 566 case subjects in those studies are not included in this analysis. In the Women's Health Initiative (N = 190 case subjects), an inverse association with α -carotene was suggested with breast cancer overall (top vs bottom tertile RR = 0.75, 95% CI = 0.49 to 1.15, P_{trend} = .19) and statistically significant among invasive tumors (RR = 0.55, 95% CI = 0.34 to 0.90, P_{trend} = .02) and case subjects diagnosed less than 5 years after blood draw (eg, 1–3 years: RR = 0.42, 95 % CI = 0.23 to 0.75, P_{trend} = .002) (35). In contrast with our results, a suggestive positive association was observed with lycopene among invasive tumors (RR = 1.47, 95% CI = 0.98 to 2.22, P_{trend} = .06). In E3N–EPIC (N = 366 case subjects), no statistically significantly associations with breast cancer risk were observed; however, many of the results were largely consistent with our results (eg, total carotenoids: top vs bottom quintile RR = 0.74, 95% CI = 0.47 to 1.16, P_{trend} = .38) (34). Thus, it is likely that our pooled results would not have been substantially altered had we been able to include these two additional studies.

A unique advantage of this analysis was the ability to accommodate true population differences in carotenoid levels between studies by recalibrating previously assayed blood samples to a common standard. Results were fairly consistent and robust regardless of whether we pooled study-specific relative risks or calculated relative risks from pooled recalibrated data. However, there was statistically significant between-study heterogeneity with original data and study-specific quintiles that was not observed with recalibrated data. Given that population differences in individual carotenoid levels were evident with recalibration, heterogeneity using study-specific quintiles was not unexpected.

Another strength of this analysis was the ability to examine associations by ER status. Given that ER⁻ tumors are less common, there was limited statistical power within each individual study to study these tumors. The inverse associations we observed among ER⁻ tumors highlight carotenoids as one of the first modifiable risk factors for this poor prognosis tumor type. Most wellestablished breast cancer risk factors are hormonal factors that are more strongly associated with ER⁺ tumors, such as parity, age at first birth, and postmenopausal BMI (47,48). Although experimental evidence has shown some carotenoids to inhibit growth of both ER+ and ER⁻ breast cancer cell lines (11), it is possible that an effect of carotenoids on ER⁺ tumors is masked by the hormonerelated associations that dominate as risk factors for ER⁺ tumors.

Recent studies of dietary intake and breast cancer risk by ER status support our findings. An analysis of the Dietary Approaches to Stop Hypertension diet in the NHS found that a higher intake of fruits and vegetables was inversely associated with ER-, but not ER⁺, tumors (top vs bottom quintile RR = 0.71, 95% CI= 0.55 to 0.90, $P_{\text{trend}} = .005$) (49). Similar associations with dietary patterns reflecting higher fruit and/or vegetable consumption also were observed in the Melbourne Collaborative Cohort Study (16), the Black Women's Health Study (17,18), and the NIH-AARP study (19). In contrast, in the E3N-EPIC study, the healthy/ Mediterranean dietary pattern was inversely associated with ER+/ progesterone receptor negative tumors (top vs bottom quartile RR = 0.65, 95% CI = 0.49 to $0.87, P_{trend} = .001$) (20). Most recently, in pooled analyses of 18 cohort studies, intakes of both fruits and vegetables (S Y Jung, D Spiegelman, L Baglietto, et al, unpublished observations) and carotenoids specifically (21) were inversely associated with ER- tumors. Statistically significant differences in the associations between carotenoids and breast cancer by ER status were observed for α -carotene, β -carotene, β -cryptoxanthin, and lutein+zeaxanthin (eg, β -carotene: ER⁻: top vs bottom quintile RR = 0.85, 95% CI = 0.77 to 0.93, $P_{\text{trend}} = .002$; ER⁺: RR = 1.04, 95% CI = 0.98 to 1.09, $P_{\text{trend}} = .24$; $P_{\text{heterogeneity}} = .01$) (21).

Given the potential antioxidant effects of carotenoids, it is possible that women who smoke or consume alcohol, lifestyle factors associated with oxidative stress, may gain more benefit from carotenoids. We observed statistically significantly stronger inverse associations of lutein+zeaxanthin and total carotenoids with breast cancer risk among current smokers, and similar patterns were observed with the other carotenoids. We did not observe statistically significant differences by alcohol intake. Interestingly, although obesity also contributes to oxidative stress (50,51), we observed interactions with BMI in the opposite direction, with statistically significantly stronger inverse associations with carotenoids among leaner women and statistically significant positive associations among obese women. This finding was unexpected and not easily explained. BMI is inversely associated with plasma carotenoid levels (3), as observed in our data, as well as adipose tissue carotenoid levels (52). However, obese women with high levels of plasma carotenoids likely also have very high adipose tissue levels. Although carotenoid levels in breast adipose tissue have been inversely associated with breast cancer risk (53), very high concentrations of carotenoids have been shown to act as pro-oxidants in animal and in vitro studies, although it is unclear whether such prooxidant activities occur in humans (54-57).

There are several proposed mechanisms by which carotenoids may influence carcinogenesis. α -carotene, β -carotene, and β -cryptoxanthin may decrease cancer risk indirectly through their metabolism to retinol, which in turn regulates cell growth, differentiation, and apoptosis via direct and indirect effects on gene expression (10,58,59). Carotenoids also may be directly anticarcinogenic by several other mechanisms, including improved gapjunction communication, enhanced immune system functioning, or antioxidant scavenging of reactive oxygen species (5–9,60–62); this may inhibit cellular dysregulation or DNA damage.

There are several limitations to our study. First, despite biologic plausibility that carotenoids are responsible for the observed inverse associations with breast cancer risk, there are alternative explanations for our findings. Other phytochemicals in fruits and vegetables, such as flavonoids and other phenolic compounds, or an interaction among various phytochemicals (63) could be associated with carotenoids and responsible for the observed association. Future studies should explore the associations between additional phytochemical components and breast cancer risk. There also is the possibility of unmeasured confounding. However, associations persisted after adjustment for other health-related behaviors, including physical activity and postmenopausal hormone use, as well as exclusion of people with health-associated behaviors, such as multivitamin use. Second, for this analysis only one blood sample was available from each participant. However, reproducibility of circulating carotenoids over a few years was very good in two of the included cohorts; intraclass correlation coefficients for individual carotenoids measured over a 2 to 3 year period were 0.63 to 0.85 in New York University Women's Health Study (28) and 0.73 to 0.88 in NHS (64). Third, although circulating carotenoids are indirect markers of activity at the breast tissue, plasma levels are positively correlated with breast tissue levels (r = 0.44-0.88) (65). Given the relatively high correlations among individual carotenoids, isolating the independent effects of a specific carotenoid is difficult. However, the inverse association with lycopene persisted with adjustment for other carotenoids, despite its correlations with other carotenoids (eg, r = 0.25 with β -carotene, r = 0.42with α -carotene). Finally, although we conducted a large number of analyses, our hypotheses were biologically motivated, and in some cases, have been published previously in the individual studies included in the pooled analysis. However, we interpret the results with caution and note that future studies should be conducted to confirm our findings.

The results of this large pooled analysis suggest that women with higher circulating carotenoid levels are at reduced breast cancer risk. The statistically significant positive associations between circulating carotenoids and risk we observed among overweight and obese women warrant further study. Additional work also is needed to determine if carotenoids are the causal factor in the observed associations. Given the possibility that another bioactive compound is responsible for the observed associations, as well as the uncertainty about the specific carotenoid(s) that are important, use of specific carotenoid supplements is not advised and may indeed be harmful among smokers (66,67). Carotenoids are available in a wide variety of fruits and vegetables common to the US diet, including: carrots for α -carotene; sweet potatoes and leafy greens for β -carotene; citrus fruits for β -cryptoxanthin; leafy greens for lutein+zeaxanthin; and tomatoes for lycopene (68). A diet high in carotenoid-rich fruits and vegetables offers many health benefits, including a possible reduced risk of breast cancer.

References

- 1. Namitha KK, Negi PS. Chemistry and biotechnology of carotenoids. Crit Rev Food Sci Nutr. 2010;50(8):728–760.
- Motchnik PA, Frei B, Ames BN. Measurement of antioxidants in human blood plasma. *Meth Enzymol.* 1994;234:269–279.
- Willett W. Nutritional Epidemiology. New York: Oxford University Press; 1998.
- Krinsky NI. Carotenoids and cancer: basic research studies. In Frei B, ed. *Natural Antioxidants in Human Health and Disease*. New York: Academic Press; 1994:239–261.
- Bertram JS. Dietary carotenoids, connexins and cancer: what is the connection? *Biochem Soc Trans*. 2004;32(Pt 6):985–989.
- Bendich A. Carotenoids and the immune response. J Nutr. 1989;119(1): 112–115.
- Krinsky NI. Actions of carotenoids in biological systems. Annu Rev Nutr. 1993;13:561–587.
- Krinsky NI. Antioxidant functions of carotenoids. Free Radic Biol Med. 1989;7(6):617–635.
- Shultz TD, Chew BP, Seaman WR, Luedecke LO. Inhibitory effect of conjugated dienoic derivatives of linoleic acid and beta-carotene on the in vitro growth of human cancer cells. *Cancer Lett.* 1992;63(2):125–133.
- Sporn MB, Roberts AB. Role of retinoids in differentiation and carcinogenesis. *Cancer Res.* 1983;43(7):3034–3040.
- Prakash P, Russell RM, Krinsky NI. In vitro inhibition of proliferation of estrogen-dependent and estrogen-independent human breast cancer cells treated with carotenoids or retinoids. *J Nutr.* 2001;131(5):1574–1580.
- Smith-Warner SA, Spiegelman D, Yaun SS, et al. Intake of fruits and vegetables and risk of breast cancer: a pooled analysis of cohort studies. *JAMA*. 2001;285(6):769–776.
- Prentice RL, Caan B, Chlebowski RT, et al. Low-fat dietary pattern and risk of invasive breast cancer: the Women's Health Initiative Randomized Controlled Dietary Modification Trial. JAMA. 2006;295(6):629–642.

- Hu F, Wang Yi B, Zhang W, et al. Carotenoids and breast cancer risk: a metaanalysis and meta-regression. *Breast Cancer Res Treat*. 2012;131(1):239–253.
- Fung TT, Hu FB, McCullough ML, Newby PK, Willett WC, Holmes MD. Diet quality is associated with the risk of estrogen receptor-negative breast cancer in postmenopausal women. *J Nutr.* 2006;136(2):466–472.
- Baglietto L, Krishnan K, Severi G, et al. Dietary patterns and risk of breast cancer. Br J Cancer. 2011;104(3):524–531.
- Boggs DA, Palmer JR, Wise LA, et al. Fruit and vegetable intake in relation to risk of breast cancer in the Black Women's Health Study. *Am J Epidemiol.* 2010;172(11):1268–1279.
- Agurs-Collins T, Rosenberg L, Makambi K, Palmer JR, Adams-Campbell L. Dietary patterns and breast cancer risk in women participating in the Black Women's Health Study. *Am J Clin Nutr.* 2009;90(3):621–628.
- Park Y, Brinton LA, Subar AF, Hollenbeck A, Schatzkin A. Dietary fiber intake and risk of breast cancer in postmenopausal women: the National Institutes of Health–AARP Diet and Health Study. *Am J Clin Nutr.* 2009;90(3):664–671.
- Cottet V, Touvier M, Fournier A, et al. Postmenopausal breast cancer risk and dietary patterns in the E3N–EPIC prospective cohort study. *Am J Epidemiol.* 2009;170(10):1257–1267.
- Zhang X, Spiegelman D, Baglietto L, et al. Carotenoid intakes and risk of breast cancer defined by estrogen receptor and progesterone receptor status: a pooled analysis of 18 prospective cohort studies. *Am J Clin Nutr.* 2012;95(3):713–725.
- Mangels AR, Holden JM, Beecher GR, Forman MR, Lanza E. Carotenoid content of fruits and vegetables: an evaluation of analytic data. *J Am Diet* Assoc. 1993;93(3):284–296.
- Micozzi MS, Beecher GR, Taylor PR, Khachik F. Carotenoid analyses of selected raw and cooked foods associated with a lower risk for cancer. J Natl Cancer Inst. 1990;82(4):282–285.
- Romieu I, Parra S, Hernández JF, Madrigal H, Willett W, Hernández M. Questionnaire assessment of antioxidants and retinol intakes in Mexican women. *Arch Med Res.* 1999;30(3):224–239.
- Eliassen AH, Colditz GA, Peterson KE, et al. Biomarker validation of dietary intervention in two multiethnic populations. *Prev Chronic Dis.* 2006;3(2):A44.
- 26. Dorgan JF, Sowell A, Swanson CA, et al. Relationships of serum carotenoids, retinol, alpha-tocopherol, and selenium with breast cancer risk: results from a prospective study in Columbia, Missouri (United States). *Cancer Causes Control.* 1998;9(1):89–97.
- Hultén K, Van Kappel AL, Winkvist A, et al. Carotenoids, alpha-tocopherols, and retinol in plasma and breast cancer risk in northern Sweden. *Cancer Causes Control.* 2001;12(6):529–537.
- Toniolo P, Van Kappel AL, Akhmedkhanov A, et al. Serum carotenoids and breast cancer. Am J Epidemiol. 2001;153(12):1142–1147.
- Sato R, Helzlsouer KJ, Alberg AJ, Hoffman SC, Norkus EP, Comstock GW. Prospective study of carotenoids, tocopherols, and retinoid concentrations and the risk of breast cancer. *Cancer Epidemiol Biomarkers Prev.* 2002;11(5):451–457.
- Tamimi RM, Hankinson SE, Campos H, et al. Plasma carotenoids, retinol, and tocopherols and risk of breast cancer. Am J Epidemiol. 2005;161(2):153–160.
- Sesso HD, Buring JE, Zhang SM, Norkus EP, Gaziano JM. Dietary and plasma lycopene and the risk of breast cancer. *Cancer Epidemiol Biomarkers Prev.* 2005;14(5):1074–1081.
- 32. Dorjgochoo T, Gao Y-T, Chow W-H, et al. Plasma carotenoids, tocopherols, retinol and breast cancer risk: results from the Shanghai Women Health Study (SWHS). *Breast Cancer Res Treat*. 2009;117(2):381–389.
- 33. Epplein M, Shvetsov YB, Wilkens LR, et al. Plasma carotenoids, retinol, and tocopherols and postmenopausal breast cancer risk in the Multiethnic Cohort Study: a nested case–control study. *Breast Cancer Res*. 2009;11(4):R49.
- Maillard V, Kuriki K, Lefebvre B, et al. Serum carotenoid, tocopherol and retinol concentrations and breast cancer risk in the E3N–EPIC study. Int J Cancer. 2010;127(5):1188–1196.
- 35. Kabat GC, Kim M, Adams-Campbell LL, et al. Longitudinal study of serum carotenoid, retinol, and tocopherol concentrations in relation to breast cancer risk among postmenopausal women. *Am J Clin Nutr.* 2009;90(1): 162–169.

1914 Articles | JNCI

- 36. Willett WC, Polk BF, Underwood BA, et al. Relation of serum vitamins A and E and carotenoids to the risk of cancer. N Engl J Med. 1984;310(7): 430–434.
- 37. Knekt P, Aromaa A, Maatela J, et al. Serum vitamin A and subsequent risk of cancer: cancer incidence follow-up of the Finnish Mobile Clinic Health Examination Survey. Am J Epidemiol. 1990;132(5):857–870.
- Wald NJ, Boreham J, Hayward JL, Bulbrook RD. Plasma retinol, betacarotene and vitamin E levels in relation to the future risk of breast cancer. *Br J Cancer*. 1984;49(3):321–324.
- Wald NJ, Nicolaides-Bouman A, Hudson GA. Plasma retinol, beta-carotene and vitamin E levels in relation to the future risk of breast cancer. Br *J Cancer*. 1988;57(2):235.
- Comstock GW, Helzlsouer KJ, Bush TL. Prediagnostic serum levels of carotenoids and vitamin E as related to subsequent cancer in Washington County, Maryland. *Am J Clin Nutr.* 1991;53(1 Suppl):260S–264S.
- El-Sohemy A, Baylin A, Kabagambe E, Ascherio A, Spiegelman D, Campos H. Individual carotenoid concentrations in adipose tissue and plasma as biomarkers of dietary intake. *Am J Clin Nutr.* 2002;76(1):172–179.
- Rosner B. Percentage points for a generalized ESD many-outlier procedure. *Technometrics*. 1983;25(2):165–172.
- DerSimonian R, Laird N. Meta-analysis in clinical trials. Control Clin Trials. 1986;7(3):177–188.
- 44. Smith-Warner SA, Spiegelman D, Ritz J, et al. Methods for pooling results of epidemiologic studies: the Pooling Project of Prospective Studies of Diet and Cancer. Am J Epidemiol. 2006;163(11):1053–1064.
- Durrleman S, Simon R. Flexible regression models with cubic splines. Stat Med. 1989;8(5):551–561.
- Marshall RJ, Chisholm EM. Hypothesis testing in the polychotomous logistic model with an application to detecting gastrointestinal cancer. *Stat Med.* 1985;4(3):337–344.
- Colditz GA, Rosner BA, Chen WY, Holmes MD, Hankinson SE. Risk factors for breast cancer according to estrogen and progesterone receptor status. *J Natl Cancer Inst.* 2004;96(3):218–228.
- Althuis MD, Fergenbaum JH, García-Closas M, Brinton LA, Madigan MP, Sherman ME. Etiology of hormone receptor-defined breast cancer: a systematic review of the literature. *Cancer Epidemiol Biomarkers Prev.* 2004;13(10):1558–1568.
- 49. Fung TT, Hu FB, Hankinson SE, Willett WC, Holmes MD. Lowcarbohydrate diets, dietary approaches to stop hypertension-style diets, and the risk of postmenopausal breast cancer. *Am J Epidemiol.* 2011;174(6): 652–660.
- Keaney JF, Larson MG, Vasan RS, et al. Obesity and systemic oxidative stress: clinical correlates of oxidative stress in the Framingham Study. *Arterioscler Thromb Vasc Biol.* 2003;23(3):434–439.
- Pou KM, Massaro JM, Hoffmann U, et al. Visceral and subcutaneous adipose tissue volumes are cross-sectionally related to markers of inflammation and oxidative stress: the Framingham Heart Study. *Circulation*. 2007;116(11):1234–1241.
- Virtanen SM, van't Veer P, Kok F, Kardinaal AF, Aro A. Predictors of adipose tissue carotenoid and retinol levels in nine countries. The EURAMIC Study. Am J Epidemiol. 1996;144(10):968–979.
- 53. Zhang S, Tang G, Russell RM, et al. Measurement of retinoids and carotenoids in breast adipose tissue and a comparison of concentrations in breast cancer cases and control subjects. *Am J Clin Nutr.* 1997;66(3):626–632.
- 54. Amengual J, Lobo GP, Golczak M, et al. A mitochondrial enzyme degrades carotenoids and protects against oxidative stress. *FASEB J*. 2011;25(3):948–959.
- Lobo GP, Amengual J, Palczewski G, Babino D, Lintig von J. Mammalian carotenoid-oxygenases: key players for carotenoid function and homeostasis. *Biochim Biophys Acta*. 2012;1821(1):78–87.
- Young AJ, Lowe GM. Antioxidant and prooxidant properties of carotenoids. Arch Med Res. 2001;385(1):20–27.
- Lowe GM, Vlismas K, Young AJ. Carotenoids as prooxidants? Mol Aspects Med. 2003;24(6):363–369.
- Gudas L, Sporn M, Roberts, AB. Cellular biology and biochemistry of the retinoids. In: Sporn M, Roberts AB, Goodman DS, eds. *The Retinoids: Biology, Chemistry and Medicine*. New York: Raven Press; 1994:443–520.

- Simeone A-M, Tari AM. How retinoids regulate breast cancer cell proliferation and apoptosis. *Cell Mol Life Sci.* 2004;61(12):1475–1484.
- Fornelli F, Leone A, Verdesca I, Minervini F, Zacheo G. The influence of lycopene on the proliferation of human breast cell line (MCF-7). *Toxicol In Vitro*. 2007;21(2):217–223.
- Sies H, Stahl W. Vitamins E and C, beta-carotene, and other carotenoids as antioxidants. Am 7 Clin Nutr. 1995;62(6 Suppl):1315S-1321S.
- Frei B. Reactive oxygen species and antioxidant vitamins: mechanisms of action. Am J Med. 1994;97(3A):5S–13S; discussion22S–28S.
- Liu RH. Health benefits of fruit and vegetables are from additive and synergistic combinations of phytochemicals. *Am J Clin Nutr.* 2003;78(3 Suppl):517S–520S.
- 64. Kotsopoulos J, Tworoger SS, Campos H, et al. Reproducibility of plasma and urine biomarkers among premenopausal and postmenopausal women from the Nurses' Health Studies. *Cancer Epidemiol Biomarkers Prev.* 2010;19(4):938–946.
- 65. Yeum KJ, Ahn SH, Rupp de Paiva SA, Lee-Kim YC, Krinsky NI, Russell RM. Correlation between carotenoid concentrations in serum and normal breast adipose tissue of women with benign breast tumor or breast cancer. *J Nutr.* 1998;128(11):1920–1926.
- Omenn GS, Goodman GE, Thornquist MD, et al. Effects of a combination of beta carotene and vitamin A on lung cancer and cardiovascular disease. N Engl J Med. 1996;334(18):1150–1155.
- 67. The effect of vitamin E and beta carotene on the incidence of lung cancer and other cancers in male smokers. The Alpha-Tocopherol, Beta Carotene Cancer Prevention Study Group. N Engl J Med. 1994;30(15): 1029–1035.
- 68. US Department of Agriculture, Agricultural Research Service. 2011. USDA National Nutrient Database for Standard Reference, Release 24. http://www.ars.usda.gov/nutrientdata. Accessed October 15, 2012.

Funding

This work was supported primarily by the National Cancer Institute at the National Institutes of Health (R03 CA128073 to AHE). Funding to individual cohorts included grants from the National Cancer Institute (R01 CA131218, P01 CA87969, R01 CA49449, R01 CA098661, P30 CA016087, P01 CA33619, R37 CA54281, CA-047988, R01 CA106591, R37 CA70867); the National Institute of Environmental Health Sciences Center (ES000260); the National Heart Lung and Blood Institute (HL043851, HL080467, HL099355); contracts from the National Institutes of Health (N01 PC35137, N01 PC35139, N02 CP1101066); the Intramural Research Program of the National Institutes of Health; the Swedish Cancer Society; Swedish Scientific Council; Swedish Research Council; Regional Government of Västerbotten, Sweden; and a grant from DSM Nutritional Products, Inc. (formerly Roche Vitamins). SJH was supported in part by a training grant (5 T32 CA09001-35) from the National Institutes of Health.

Notes

The study sponsors had no role in the design of the study; the collection, analysis, and interpretation of the data; the writing of the manuscript; or the decision to submit the manuscript for publication.

Affiliations of authors: Channing Division of Network Medicine (AHE, RMT, SEH) and Division of Preventive Medicine (JEB, HDS), Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, MA; Department of Epidemiology (AHE, SJH, RMT, SEH) and Department of Nutrition (SJH, HC), Harvard School of Public Health, Boston, MA; Division of Cancer Epidemiology and Genetics, National Cancer Institute, National Institutes of Health, Rockville, MD (LAB); Vanderbilt University School of Medicine, Nashville, TN (QD, WZ); Fox Chase Cancer Center, Philadelphia, PA (JFD); University of Hawaii Cancer Center, Honolulu, HI (AAF, MTG, LRW); Shanghai Cancer Institute, Shanghai, China (Y-tG); Department of Public Health and Clinical Medicine, Umeå University, Umeå, Sweden (GH, KH); George W. Comstock Center for Public Health Research and Prevention, Johns Hopkins University, Bloomberg School of Public Health, Baltimore, MD (KJH, JH-B); The Prevention and Research Center, Mercy Medical Center, Baltimore, MD (KJH); Division of Health Studies, Agency for Toxic Substances and Disease Registry, Centers for Disease Control and Prevention, Atlanta, GA (ALS); Department of Obstetrics and Gynecology (PT) and Department of Environmental Medicine (AZ-J), New York University Cancer Institute, New York University School of Medicine, New York, NY; Department of Clinical Nutrition, University of Gothenburg, Göteborg, Sweden (AW); Division of Biostatistics and Epidemiology, School of Public Health and Health Sciences, University of Massachusetts, Amherst, MA (SEH).