



Treatment of lymphomas via regulating the Signal transduction pathways by natural therapeutic approaches: A review

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ABSTRACT

Lymphoma is a heterogeneous group of malignancies, which comprises 4.2 % of all new cancer cases and 3.3 % of all cancer deaths in 2019, globally. The dysregulation of immune system, certain bacterial or viral infections, autoimmune diseases, and immune suppression are associated with a high risk of lymphoma. Although several conventional strategies have improved during the past few decades, but their detrimental impacts remain an obstacle to be resolved. However, natural compounds are considered a good option in the treatment of lymphomas because of their easy accessibility, specific mode of action, high biodegradability, and cost-effectiveness. Vegetables, fruits, and beverages are the primary sources of natural active compounds. The present review investigated the activities of different natural medicinal compounds including curcumin, MK615, resveratrol, bromelain, EGCG, and *Annonaceous acetogenins* to treat lymphomas. Moreover, *in vitro* and *in vivo* studies, classification, risk factors, and diagnosis of lymphoma are also discussed in the present review. The accumulated data proposed that natural compounds regulate the signaling pathways at the level of cell proliferation, apoptosis, and cell cycle to exhibit anti-lymphoma activities both *in-vivo* and *in-vitro* studies and suggested that these active compounds could be a good therapeutic option in the treatment of different types of lymphomas.

1. Introduction

Lymphomas are known as a heterogeneous group of malignancies, with multiple sets of causes, natural progression and comprise more than 3 % of all malignancies [1,2]. It is a cancer of the lymphatic system, which acts as a filter of the body consist of lymph nodes and a web of vessels [3]. All lymphoid malignancies originate from systemic lymphoid tissues and can spread over the other parts of the body [4]. According to accumulated data, approximately 85–90 % of lymphomas are derived from B lymphocyte cells, whereas remaining originate from T lymphocyte cells and natural killer cells [5]. Lymphomas are majorly classified into two main classes including Non- Hodgkin lymphoma (NHL) (accounts 90 % of lymphomas) and Hodgkin's lymphoma (HL)

(accounts for only 10 % of all lymphomas) [5]. Lymphoid malignancies are presented with highly diverse features including enlarged lymph nodes and systemic symptoms of fatigue, fever, weight loss, and extra lymphatic spread are also seen initially in patients who are suffering from both HL and NHL [6,7].

According to the emerging evidence, the NHL is ranked as the 7th most general cancer among men and 6th most commonly diagnosed cancer among women [8]. Globally, approximately 510,000 new confirmed cases of NHL were estimated in 2018, and wildly it is ranked as the 5th - 9th most common cancer in most of the countries [9]. The incidences and deaths due to NHL across the globe vary with different geographical regions [10]. In most of the Middle East countries, the relative frequency of NHL seems to be lower than HL in contrast with the

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global burden of diseases. A study reported that the incidence rate of NHL was higher among developing countries including Pakistan, India, Sudan, and Bangladesh which comprise about 50.5 % of the world cases. Globally, a 2.4 % death rate was estimated due to NHL and two-third of them (125,000) deaths were occurring in underdeveloped countries due to poor survival in these regions [11]. The emerging evidence suggested that the Burkitt lymphoma (BL), diffuse large B cell lymphoma, anaplastic lymphoma and lymphoblastic lymphoma (LL) are major types identified in children mostly [12]. The incidence rate of NHL was higher specifically Burkitt lymphoma and HL which comprises 6 and 3% respectively in children, whereas HL accounts for 13 % of adolescent cancers (aged 15–19 years) [13].

According to emerging evidence, the dysregulation of the immune system is considered as one of the high-risk factors for lymphoma [14]. The autoimmune disorder is also associated with the best-established risk factor for lymphomas. In addition, the few medications (tumor necrosis inhibitors) during the treatment of autoimmune disorder are also associated with a high risk of Hodgkin and non-Hodgkin Lymphomas [15]. Younes and co-workers revealed that the occurrence of lymphomas also associated with a bacterial and viral infection like Epstein-Barr virus caused the risk of high-grade B-cell, primary effusion, and Burkitt lymphomas [16]. However, the diagnosis is a necessary and important thing before the treatment of distinct types of lymphomas. For diagnostic evaluation, the determination of tissue is required to check the presence of lymphoma for the treatment plan [17]. Needle core biopsy is increasingly used for lymphadenopathy malignancy diagnosis, it not only provides a definite diagnosis but also helped in the differentiation of multiple lymphomas [18]. In addition to biopsy, biomarkers including an elevated level of serum cytokines [19] and lactate dehydrogenase (LDH) are also used for the diagnosis of lymphomas [20].

According to accumulated data, different therapies including chemotherapy (dacarbazine, bleomycin, or doxorubicin) and radiotherapy are considered as standard single or combined treatment modalities against lymphomas [16]. A recently published study reported that in addition to the positive effects of these therapies, they also exert a number of adverse effects on different body organs including skin, reproductive, cardiovascular, renal, hematological, and neurological systems [21]. Human epidemiological studies showed that the consumption of natural products is considered a promising therapy against different cancers including lymphomas due to their less adverse effects, specificity, high biodegradability, easy accessibility, and greater mode of action [22–24]. The *in-vivo* and *in-vitro* studies reported that the medicinal plants (*Glycine max*, *Ananas Comosus*, *Vitis vinifera*, *Prunus mume*, *Curcuma longa*, *Momordica charantia*, and *Zingiber Officinale*) modulate different signal pathways including apoptosis, inhibition of cell proliferation, cell cycle arrest at (G1/S phase), deactivation/activation of NF- κ B pathway, regulation of signaling caspases, up-regulation of antioxidant enzymes, epigenetic modulation, metastasis, and inhibition of PI3K/Akt/mTOR pathway in the treatment of lymphomas [14, 25–29]. Shanmugam et al. [30] reported that curcumin (natural compound) induced apoptosis and cell-cycle arrest (G1/S) in the treatment of Hodgkin lymphoma. Similarly, 25–300 μ M administration of resveratrol effectively regulates the death receptor pathway by activation of EBV lytic antigens during the treatment of Raji/Akata cell lines of lymphocytes [31].

However, according to the epidemiology of lymphoma, there is a need to develop a strong therapeutic drug with unique characteristics and less adverse effects to control the mortality rate. To handle this situation, despite research manuscripts, the review article has equal importance to highlight and recommend the solutions. The present review investigates the anti-lymphomas activities of *Curcuma longa*, *Prunus mume*, *Vaccinium macrocarpon* (resveratrol), *Ananas comosus*, *Camellia sinensis*, and *Annona muricata*. Moreover, the classification, risk factors, and diagnosis of lymphomas are also discussed in the present review for a better understanding.

Table 1

The classification of Hodgkin and non-Hodgkin lymphoma.

Main class	Subclass	Percentage (%)	Reference
Hodgkin's Lymphoma	Nodular sclerosis classical Hodgkin's lymphoma (NSCHL)	60–75	[38,112]
	Mixed cellularity classical Hodgkin's lymphoma (MCCHL)	20–25	[38,112]
	Lymphocyte-depleted classical Hodgkin's lymphoma (LDCHL)	less than 1	[38,112]
	Lymphocyte-rich classical Hodgkin's lymphoma (LRCHL)	5 with LRCHL	[38,112]
	Nodular Lymphocyte-predominant HL (NLPHL)	5	[112, 113]
	Diffused large B- cell lymphoma (DLBCL)	32.5	[114]
	Follicular Lymphoma (FL)	17.1	[114]
	Marginal Zone lymphoma (MZL)	8.3	[114, 115]
	Mantle-cell lymphoma (MCL)	3–10	[116]
	Peripheral T-cell lymphoma (not otherwise specified – PTCL (NOS))	1.7	[114]
	Non-Hodgkin's Lymphomas	Burkitt lymphoma (BL)	1.6–2.2
Anaplastic large cell lymphoma (ALCL)		1	[114]
Extranodal NK/T-cell lymphoma (ENKL)		2.2	[115]
Mucosa-associated lymphoid-tissue (MALT) lymphoma		5	[117]
Lymphoplasmacytic lymphoma (LPL)		1.1	[114, 118]
Others		18.9	[114]

2. Classification of lymphoma

Lymphomas are classified into two main classes including rare Hodgkin and common non- Hodgkin lymphoma [32]. Nodular lymphocyte-predominant Hodgkin lymphoma (LPHL) is a relatively uncommon HL that comprises 5–10 % of all Hodgkin lymphomas and shows unique clinicopathological features (cervical region and axillary region) compared to classical HL [33]. Classical HL account for about 85 % of all HL cases [33]. Nodular sclerosis classical HL (NSCHL) [34], mixed cellularity HL [35], lymphocyte-depleted HL (LDCHL) [36] and lymphocyte- rich classical Hodgkin's lymphoma (LRCHL) are further subgroups of classical Hodgkin's lymphoma [37,38], as presented in Table 1. NSCHL exhibits major differences from lymphocyte-depleted (LD) and mixed cellularity (MC) HL, so these are recognized as separate entities [33]. More common lymphoma which occurred all over the world is NHL, there are many subtypes of NHL as presented in Table 1. DLBCL is the most common subclass of NHL in elderly people [39], and FL is considered as the second most common subtype of NHL in the Western population [40]. According to accumulated data, yet more than 50 various subtypes of T and B-cell NHL had been defined, each subtype has unique clinic-pathological and biological characteristic [41].

3. Risk factors for lymphoma

Cancer incidences and deaths vary in different geographical regions because of unequal distribution of risk factors for cancer [10]. Regarding the entity of risk factors of lymphoma, extensive investigation is done to govern how and whether lymphomas are associated with prior medical conditions, fitness records, or other conditions including blood transfusions, smoking, or use of other carcinogens. Few risk factors of lymphomas are presented in the following sections.

3.1. Genetics

Cytokines are small signaling proteins that are produced and released primarily by cells of innate and adaptive immunity, but also by

epithelial and stromal cells and help in the regulation of immune responses to external stimuli (temperature, heat, environmental factors) [42,43]. Cytokine proteins play an important role in immune regulation [44]. The key function of cytokines is to regulate the inflammatory response. The disturbed balance between immune response and anti-tumor activity leads to cancer development [44]. Nuclear factor-kappa B (NF- κ B) and JAK/STAT pathways are reported as the signaling pathways for cytokines' receptors [45,46]. Tumor burden in patients is reflected by serum specific cytokine profiles. The soluble mediators' cytokines are produced by reactive cells or tumor cells which can contribute to tumor development [47]. Tumor necrosis factor (TNF) and Interleukin 10 (IL10) are the genes that code for some immune-regulatory cytokines which are associated with serious mediators of programmed cell death, inflammation and T-cell subsets (Type 1 T-helper cell and Type 2 T-helper cell balance), and function as autocrine growth factors (HB-EGF and Amphiregulin) in lymphoma [48–50].

3.2. Polymorphism in xenobiotic metabolizing enzyme

Polymorphism in genes that encoded xenobiotic metabolism enzymes (N-acetyltransferases and glutathione S-transferase) increased the susceptibility of developing lymphomagenesis [51,52]. A study suggested that the people who mostly use hair dyes are at higher risk of NHL, especially females. Chemicals present in hair dyes cause polymorphisms that disturbed the enzymic activity and its normal functioning [53].

3.3. Acquired and autoimmune situations

Certain acquired and autoimmune situations, hypersensitive reactions, and inflammatory diseases such as primary Sjogren's syndrome (pSS), rheumatoid arthritis, and systemic lupus erythematosus (SLE), are considered as high-risk factors for lymphoma [5,16,54]. Several studies conducted in different patients showed that the high incidences of lymphoma are mainly associated with an autoimmune condition. But the relationship between lymphoma and autoimmune condition remains unclear [55]. Kaulen et al. [56] reported that the administration of immunosuppressive agents (azathioprine, cyclosporine, leflunomide, and chlorambucil) for autoimmune diseases (AID) and solid organ transplantations (SOT) are also considered as a high-risk factor for NHL.

3.4. Bacterial and viral infections

Among various types of cancers, lymphoma malignancies are first discovered that are associated with infectious agents cause by either bacteria or viruses including Human T-lymphotropic virus-1 (HTLV-1), and Epstein-Barr virus (EBV) particularly caused Burkitt lymphoma and high-grade B-cell lymphoma [16]. According to emerging evidence, the hepatitis B virus infection is also found to be associated with lymphoma malignancies. The patient who suffered from hepatitis B virus infection has a higher risk of B-cells (diffused large B cell lymphoma; DLBCL) [57]. *Helicobacter Pylori* (HP) was the first bacteria that was recognized to play a causative role in the development of human malignancies including lymphoma [58]. Evidence showed that bacterial infections caused Mucosa-associated lymphoid tissue Lymphoma (gastric MALT lymphoma) in humans [59,60].

4. Diagnosis

Lymphoma has extensively varying genetic abnormalities, immunophenotyping and clinical features [61]. Patients with lymphoma might suffer from ambiguous symptoms such as weight loss, and low-grade fever, or altered hematological laboratory values. The biopsy technique was considered as the most promising technique for the initial diagnosis of lymphoma [32]. The utilization of biopsy not only allows diagnosis but also helps to identify the subclassification of lymphoma.

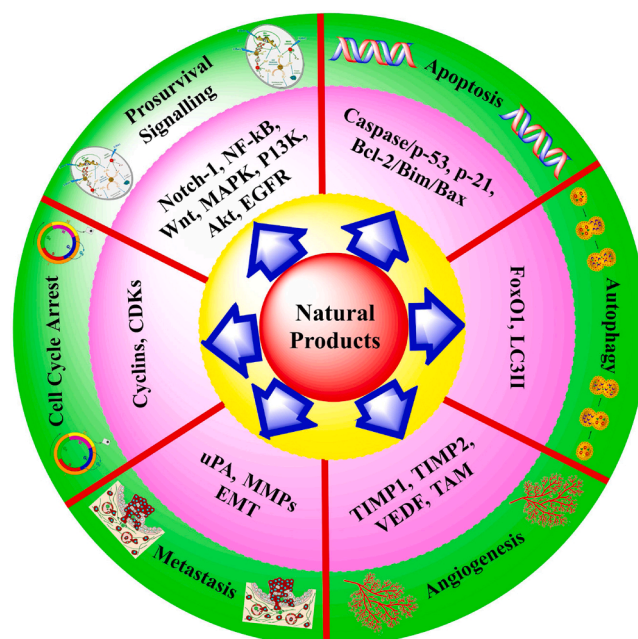


Fig. 1. The anti-cancerous activities of natural compounds by regulating different pathways.

Recently needle-core biopsy for the lymphoma diagnosis has proven as less adverse traumatic, properly tolerated, and cost-effective mainly for the patients who have deep-seated lesions or poor clinical situations [62]. A study also suggested that ultrasound-guided full core needle biopsy (UFCNB) offers the best alternatives for the diagnosis of lymphoma with a 95.0 % accuracy rate, and 97.2 and 88.7 % sensitivity for NHL and HL, respectively [63].

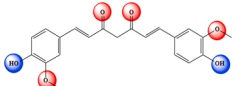
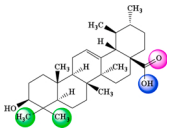
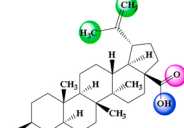

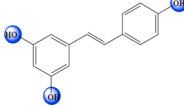
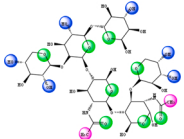
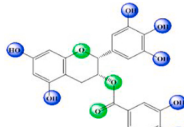
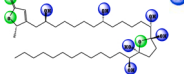
The other biochemical parameters are also used for lymphoma diagnosis. The cytokine estimation by enzyme-linked immunosorbent assays helped in the assessment of lymphoma patients at clinical and laboratory levels [64]. Besides, non-specific tumor marker namely lactate dehydrogenase (LDH) raised in the case of lymphoma [20]. Serum levels of lactate dehydrogenase are determined to diagnose lymphoma. Enzyme lactate dehydrogenase-5 catalysis the formation of lactate from pyruvate in the presence of NAD^+ at the end of the glycolysis process. The up-regulation of this process is found in several malignant tumors including lymphomas [65].

5. Medicinal plants for the treatment of lymphoma

For most lymphomas, the natural products (curcumin, MK615, resveratrol, and bromelain) are considered as a standard treatment due to their accurate mode of action, greater bioavailability, specificity, cost-effectiveness, and fewer adverse effects [21,27]. Human epidemiological statistics showed a greater connection between diets rich in vegetables, and fruits, and reduction of cardiac diseases and cancers [66]. Polyphenols are considered as potent anticancer agents, they exert their anticancer activity through a wide range of processes including they inhibit one or more biochemical pathways and biomolecules that help in the treatment of cancer cells [67,68]. One of the possible strategies for the treatment of lymphomagenesis is to target signaling pathway of NF- κ B by natural approaches that regulate the expression of thousands of genes [69]. Besides, natural compounds induce apoptosis, angiogenesis, metastatic, autophagy, cell-proliferation, and cell cycle arrest to treat the cancer cells including lymphoma (see Fig. 1) [27]. Following different medicinal plants are discussed to reduce and control the epidemiology of lymphoma.

Table 2

The representative structures of natural compounds with their sources and molecular mass.

Plant	Active compound	Molecular formula	Chemical Structure	Identification ID	Reference
Curcuma longa (Turmeric)	Curcumin	C ₂₁ H ₂₀ O ₆		969,516	[119]
	Ursolic acid	C ₃₀ H ₄₈ O ₃		64,945	[120]
Prunus mume (Japanese apricot)	Betulinic acid	C ₃₀ H ₄₈ O ₃		64,971	[121]
	Oleanolic acid	C ₃₀ H ₄₈ O ₃		10,494	[122]
Vitis vinifera (Grape vine)	Resveratrol	C ₁₄ H ₁₂ O ₃		445,154	[123]
Ananas comosus (Pineapple)	Bromelain	C ₃₉ H ₆₆ N ₂ O ₂₉		44,263,865	[124]
Camellia sinensis (Green tea)	EGCG	C ₂₂ H ₁₈ O ₁₁		65,064	[125]
Annona muricata (Annona fruits)	Annonaceous acetogenins	C ₂₆ H ₄₆ O ₇		393,472	[126]

5.1. Curcumin

Curcumin (diferuloylmethane) is a natural product of yellow-colored Phyto-polyphenol derived from *Curcuma longa* herb root (see Table 2). During the past couple of decades, curcumin extensively studied as an anti-cancerous agent against multiple myeloma including lymphomas by suppressing cell-proliferation, cellular transformation, invasion, metastasis, and angiogenesis [70,71]. Mackenzie and his research group reported that curcumin plays a significant role in the reduction/control of lymphomas of about 80.97 % by suppressing the activation of NF-κB and STAT-3 factors, cell cycle arrest, inhibit the expression of cell-proliferated proteins and apoptosis [72]. Similarly, *in vivo* study demonstrated that the nano-scaled TPGS-curcumin and SLN-curcumin effectively reduced the lymphoma (HL) effect in mice by 43 % ($p < 0.04$) and 50.5 % ($p < 0.02$) respectively. Besides, nano-scaled SLN-curcumin significantly inhibits the effect of cell-proliferation proteins including Mcl-1 and XIAP, and expression of anti-inflammatory cytokines such as TNF-α and IL-6 in the treatment of HL [73]. Guo and his research group reported that the synergistic effect of curcumin with doxorubicin showed remarkable results in the treatment of B cell lymphoma both *in vitro* and *in vivo* studies. They revealed that co-delivery of curcumin and doxorubicin (loading rate of curcumin and

doxorubicin: 8.1 and 9.7 %) into xenografted mice reduced the invasive B cell lymphoma. They also stated that curcumin and doxorubicin combination *in vitro* significantly removed the invasive B cell lymphomas by regulating the apoptosis pathways [74]. Chen et al. [75] demonstrated that curcumin inhibited the invasion, mitigation, and cell proliferation activities and promoted the apoptosis in SU-DHL-8 cell line (diffuse large B cell lymphoma) by suppression and regulating the expressions of miR-21 and Von Hippel-Lindau (VHL) respectively. Similarly, *in vivo* study reported that curcumin suppressed the regulation of NF-κB signaling pathway and expression of pro-inflammatory cytokines in the treatment of Dalton's lymphoma cells by promoting the anti-oxidative defense system [76].

Aldehyde dehydrogenase (ALDH) is considered an imported enzyme in different types of cancer [31]. Li et al. [71] stated that 1 μM dose of curcumin effectively reduced the effect of Burkitt lymphoma cell lines as BL41-2 ($p < 0.0002$) and DG-75 ($p < 0.0377$) of about 50 % by inhibiting the proliferation, whereas by increasing the dose level up-to 15 μM, the 75 % reduction of THP-1 ($p < 0.0465$) was noted in ALDH-positive population. Another study revealed that curcumin effectively suppressed the expression of TNF-α mRNA in mantle cell lymphoma. It also observed that the inhibition of TNF-α cell leads to the inactivation of NF-κB and cell-proliferation factors which were then

neutralized by TNF- α antibody [77]. However, in the primary effusion lymphoma, curcumin significantly induced apoptosis and suppressed the activity of cell-proliferation by inhibition of the STAT-3 pathway [30].

5.2. Japanese apricot (*Prunus mume*)

Prunus mume (commonly known as Japanese apricot; *Rosaceae*) is considered as a good medicinal drug and food in Japan, Korea, and China. The recent evidence suggested that *Prunus mume* possesses several biological and pharmacological activities including inhibition of influenza A virus, anti-cancerous activities, suppression of pro-inflammatory factors, induction of apoptosis, and improvement of blood fluidity [78]. The accumulated data suggested that MK615 is an important and active ingredient/extract isolated from *Prunus mume* which holds multiple cyclic triterpenes including ursolic acid, betulinic acid, oleanolic acid are known for their anti-cancerous and anti-inflammatory activities, the structures are represented in Table 2 [79]. These triterpenes are active inhibitors that released pro-inflammatory cytokines (TNF- α , IL-6, and IL-8). Cytokines inhibition is based on MK615-induced inhibition by NF- κ B pathway activation [80]. Inoue et al. [79] reported that MK615 exhibit significant anti-cancerous activity in a concentration-dependent manner against BALM-3 lymphocytic cell lines in combination with bendamustine. They also stated that combined administration of 25 μ g/mL of bendamustine with 7.5 μ g/mL of MK615 for 48 h treatment significantly showed the cytoplasmic blebbing, chromatin condensation, nuclear fragmentation, expression of Annexin-V and reduction of caspase-3 in BALM3 cells, which results in a noteworthy reduction of BALM3 cells of about 34.15 % as compared to alone MK615 treatment (1.11 %). Tsuji et al. [81] also found that *P. mume* ethanolic extract stimulates innate immunity which helps in the enhancement of immune function. Yanaki et al. [82] stated that MK615 significantly downregulated the expression of PD-L1 on tumor cells and showed a prolonged survival activity in a BL6/B16 mouse that disclosed the T cell-mediated anti-cancerous immunity. Similarly, Al-Jahdari and co-workers observed that the combination of MK615 with X-ray radiations protected CD4/CD8 T-cells and displayed the remarkable antitumor ability [83].

5.3. Resveratrol

Resveratrol (3,540-trihydroxystilbene) is belonging to non-flavonoid polyphenol class called stilbenes. It is derived from several plant and beverage sources including grapes, berries, plums, peanut, and red wines (see Table 2) [84]. According to emerging data, resveratrol possesses several properties including anticancer, anti-inflammatory, antioxidant, and antiaging activities [85]. Meng et al. [86] reported that considerable administration of resveratrol (20 μ M) effectively reduced cell-proliferation activity in HL-60 and NB-4 cells by enhancing the expression of a caspase-3 pathway. They also revealed that during the treatment of HL-60 cells, resveratrol increased the expression of PTEN mRNA and decreased the activity of p-AKT pathway. Kong et al. [87] demonstrated that the analog of resveratrol (pterostilbene) significantly induced apoptosis in the treatment of diffuse large B-cell lymphoma cell lines (DLBCL). They also reported that pterostilbene arrest the cell cycle at G1 phase in the reduction of DLBCL cell lines by inhibiting the cyclin-dependent kinase-2 (Cdk-2) and increasing the effect of checkpoint dependent kinase-2 (Chk-2). Zunino and his research group revealed the effect of resveratrol against human B lymphocytes (BL), they stated that during the treatment of B lymphocytes cell resveratrol showed dual behaviors at different concentrations. At 5 μ M, resveratrol increased the cell-proliferation of CD19⁺ BL, whereas resveratrol administration of 10 μ M significantly suppressed the cell proliferation of CD19⁺ BL and resultingly enhanced the expression of caspase-3 activation [88]. Radwan and co-workers revealed that resveratrol effectively promoted the activity of human leukocyte antigen (HLA) class II

proteins family (DM and DR) during the treatment of B cell lymphoma (BCL). They also observed that resveratrol enhanced the processing of HLA class II-mediation by altering the response of thiol reductase and endo-lysosomal cathepsins including B, D, and S during the treatment of BCL [89]. Another study conducted by Ko et al. [90] reported that resveratrol reduced the mass of anaplastic large cell lymphoma (ALCL) by regulating the expression of markers including CD8, CD3, and CD2, present on the surface of ALCL. They also observed that resveratrol significantly up-regulated the expression of death receptors like Fas during the treatment of SR-786 cell line of ALCL.

Faber and Chiles, [91] demonstrated that 25 μ M concentration of resveratrol in lymphocyte cell lines including OCI-Ly-1 and OCI-Ly-18 significantly induced the cell cycle arrest in S-phase. Another study reported that 24–32 μ M consumption of resveratrol induces apoptosis in SUDHL-4 and NU-DUL-1 cell lines by the activation of Bax/Bcl2 pathway [87]. Jara and co-workers determined the anti-proliferated effect of resveratrol on Ramos cell line of Burkitt lymphomas by estimating its cell viability through MTT and Trypan Blue exclusion assay at different concentrations after 24 and 48 h treatment respectively. Both MTT and Trypan Blue exclusion assays revealed that maximum anti-proliferative activity of resveratrol against Ramos cell line was found after 48 h treatment with 150 μ M concentration [92].

5.4. Pineapple (*Ananas comosus*)

Pineapple (*Ananas comosus*) is one of the most popular tropical fruit in the world. Pineapple is a member of the *Bromeliaceae* family [93]. Bromelain is an active protease and isolated from stem and fruit of pineapple as presented in Table 2 [94]. Bromelain exhibits several biological properties including anti-inflammatory, antibiotic, anti-cancer, anticoagulative, anti-thrombotic, and anti-edematous activities [95]. *In vitro* experiment showed that bromelain treatment to T cell eliminated the leu8/LAM1, CD8, CD44, CD6, and MIC2 molecules and promoted the expression of CD2 to activate T cell [96]. Another study demonstrated that bromelain effectively inhibited the regulation of CD25 surface molecules and blocked the expression of CD4⁺ T cells. Furthermore, bromelain inhibited the expression of ERK₂ during the activation of T-cells [97].

Debnath and co-workers studied (*in-vitro*) the effect of bromelain in combination with peroxidase (BM-PR) against Dalton's lymphoma (DLA) cells and reported that BM-PR significantly suppressed the cell-proliferation at a dose of 500 μ g/mL after 72 h treatment. They also suggested that BM-PR combination effectively inhibited the expression of an anti-apoptotic protein (Bcl-2), and enhanced the activity of apoptotic proteins including p-53, cytochrome-c, and Bax during the treatment of DLA [25]. Another study also founded that p53 expression in DLA cells was very low. But after treatment with bromelain plus peroxidase, the expression of p53 was significantly increased [26]. A similar study reported that bromelain and N-acetylcysteine proved as an important constituent in the treatment of human lymphoma cells by suppressing the expression of p-38 protein and activating the abnormal cell cycle progression [28,98]. Debnath et al. [25] reported that the administration of bromelain with peroxide (50 mg/kg; body weight) significantly reduced the activity of anti-apoptotic proteins including NF- κ B and Bcl-2, and increased the expression of pro-apoptotic proteins like p-53, Bad, and Bax against non-Hodgkin lymphoma. Debnath et al. [99] showed that bromelain (*in vivo*) treatment in combination with peroxidase significantly promoted the expression of Bax, Bad, and cytochrome c and down-regulated the expression of NF- κ B and Bcl-2 protein family during the treatment of Dalton's lymphoma cells (DLA) in mice and suggested as good therapeutic agent.

5.5. *Annona* fruits

Annona fruits belong to *Annonaceae* family that enriched with phenolic compounds such as phenolic acids, flavonoids, stilbenes,

Table 3The *in vivo* and *in vitro* studies of active compounds with a mechanism of action in the treatment of lymphomas.

Plant	Active compound	Study type	Lymphoma/cell lines	Dose	Mechanism	Reference
<i>Curcuma longa</i>	Nano-scaled TPGS-curcumin and SLN-curcumin	<i>In vivo</i>	Hodgkin's lymphoma/ L-540	10–40 μ M	Inhibit cell-proliferation and enhance the expression of anti-inflammatory cytokines (TNF- α and IL-6)	[73]
<i>Curcuma longa</i> herb root	Curcumin	<i>In vitro</i>	Cultured glioblastoma cells/ U-87MG and U-251MG	20 μ M	Inhibit PI3K/Akt/mTOR pathway	[70]
<i>Curcuma longa</i>	Curcumin	<i>In vitro</i>	Hodgkin's lymphoma cells/ KM-H2, L-428, and L-1236	25 μ M	Inhibit NF- κ B and STAT3 activation. Suppress cell proliferation and induce apoptosis	[72]
<i>Prunus mume</i> (Japanese apricot)	MK615	<i>In vitro</i>	B-lymphoma/BALM-3	7.5 μ g/mL	Synergize with bendamustine to induce apoptosis and suppress cell-proliferation	[79]
<i>Prunus mume</i>	MK615	<i>In vitro</i>	T-cell leukemia/ Su9T01 and S1T cells	45.9 μ g/mL	Inhibit cell-proliferation	[127]
<i>Vitis vinifera</i> , <i>Vaccinium macrocarpon</i>	Resveratrol	<i>In vitro</i>	Human leukemia (HL)/ HL-60	20 μ M	Inhibit cell-proliferation, increase expression of caspase-3 pathway	[86]
<i>Vaccinium macrocarpon</i>	Resveratrol (pterostilbene)	<i>In vitro</i>	B-cell lymphoma cell lines (BCL)/ SUDHL-4, NU-DUL-1, OCI-LY8, and TMD8	12.5–100 μ M	Induce apoptosis, and arrest the cell cycle at G1/S phase	[87]
<i>Vaccinium macrocarpon</i>	Resveratrol (pterostilbene)	<i>In vivo</i>	B-cell lymphoma cell lines (BCL)/OCI-LY8	30 mg/kg/20 days	Inhibits the growth of implanted OCI-LY8 cells in a xenograft mouse model	[87]
<i>Vitis vinifera</i>	Resveratrol	<i>In vitro</i>	B lymphocytes (BL)/CD19 + BL	10 μ M	Suppress the cell proliferation, and increase expression of caspase-3 pathway	[88]
<i>Annona reticulata</i>	HEEAM	<i>In vivo</i>	Dalton's lymphoma ascites (DLA)/Swiss albino mice	400 mg/kg/body weight	Decrease the PCV and arrested the tumor growth	[103]
<i>Annona muricata</i>	Phenolics, flavonoids, and tannins	<i>In vitro</i>	Lymphoma/ Ramos-1	200 μ g/mL	Suppress the cell-proliferation	[14]
<i>Annona macrophyllata</i>	Polyphenols	<i>In vivo</i>	Human leukemic monocyte lymphoma cells/ U-937	4.38 mg/kg/body weight	Synergize with geranylgeraniol to inhibit the mitochondrial complex I, and anti-proliferation	[104]
Pineapple (<i>Ananas comosus</i>)	Bromelain	<i>In vivo</i>	Leukemia/P-388	12.5 mg/kg/body weight	Trigger the action of T cytotoxic cells, and angiogenesis	[128]
Pineapple (<i>Ananas comosus</i>)	Bromelain	<i>In vivo</i>	Non-Hodgkin lymphoma (NHL)/ DLA	50 mg/kg/body weight	Synergize with peroxide to modulate the activity of apoptotic proteins, and up-regulate the antioxidant enzyme	[25]
Pineapple (<i>Ananas Comosus</i>)	Bromelain	<i>In vitro</i>	Dalton's lymphoma (DLA)	500 μ g/mL/ 72 h	Synergize with peroxidase to inhibit proliferative activity, induce apoptosis	[25,26]
<i>Camellia sinensis</i>	Green tea (epigallocatechin-3-gallate)	<i>In vivo</i>	Human leukemia (HL)/HL-60 (human xenografted SCID mice)	100 mg/kg/body weight	Synergize with quercetin to inhibit the activity of anti-apoptotic proteins (Bcl-2 and Mcl-1) and increase the expression of pro-apoptotic proteins (Bax)	[29]
<i>Camellia sinensis</i>	Epigallocatechin-3-gallate	Clinical trial	Non-Hodgkin lymphoma (NHL)	3.5 cups/day	Induce apoptosis	[111]

coumarins, tannins, and lignins. *Annona* fruits have strong antioxidant and anticancer activities because of their polyphenolic content (flavonoids, stilbenes, and coumarins) (see Table 2) [100]. The presence of phenolic compounds protect against the development of NHL because lower intakes of antioxidants linked to a compromised immune system [14]. According to the emerging evidence, *Annonaceae acetogenins* is an important active compound derived from the seeds of *Annona*, considered as a good potent therapeutic agent against several cancer cell lines [101]. A study conducted on *A. reticulata* and *A. squamosa* (species of *Annona* fruit) revealed that consumption of fruit helps in the treatment of adult T-cell leukemia/lymphoma (ATL) malignancy caused by human-cell lymphotropic virus type I (HTLV-I) [102]. Al-Shaya et al. [14] demonstrated that the chloroform extract of *Annona muricata* significantly suppressed the cell-proliferation of about 95 % in a dose-dependent manner (200 μ g/mL; $p < 0.05$) against lymphoma cell line of Ramos-1. Similarly, Nalini and Durairaj, [103] revealed that hydroethanolic extract of *Annona muricata* showed high anti-lymphoma activity at 400 mg/kg (body weight) in Dalton's lymphoma ascites (DLA)-induced tumor models in a dose-dependent manner ($p < 0.05$). Recently, a study conducted by Calzada and co-workers revealed that *Annona macrophyllata* and geranylgeraniol showed the maximum anti-lymphoma (human leukemic monocyte lymphoma cells) inhibitory concentration (EC₅₀) value of 1.07 and 4.38 mg/kg for Balb/c male mice, and 1.33 and 3.66 mg/kg for female mice respectively [104]. Similarly, Vikas et al. [105] extracted *Annona squamosa* by four different solvents (ethyl acetate, chloroform, methanol, and petroleum ether) and revealed that petroleum ether extract showed better cytotoxicity in

leukemic K-562 cells and significantly reduced the mass of Dalton's lymphoma ascites (DLA) along with breast, lung, and nasopharyngeal cancer.

5.6. Green tea (*Camellia sinensis*)

Green tea is one of the important drinks taken by Chinese and Japanese over the thousands of years and derived from the leaves of *Camellia sinensis*. Polyphenols including flavonoids are the strongest antioxidant, anti-cancer, and anti-inflammatory agents in green tea. Epicatechin gallate, epigallocatechin-3-gallate (EGCG), epigallocatechin, and catechin, are the important active compounds in *C. sinensis* (see Table 2) [106]. A study showed that green tea (catechins) effectively robust the immune response against cancer by promoting the expression of natural killer and CD4⁺ T cells. Furthermore, 4'-hydroxyl consider as a critical factor not only for promoting the activity of natural killer cells but also enhancing the expression of CD4⁺ T cells *in vivo* [107]. Another study revealed that EGCG significantly boosts the immune response against tumor cells by enhancing the quantity of CD8⁺ and CD4⁺ T cells and reducing the accommodation of myeloid-derived suppression cells (MDSC) [108].

Long-term consumption of green tea increased the activity of xenobiotic-metabolizing enzyme glutathione S-transferase (GST), which helped in the inhibition of NF- κ B transcription factors, and provide protection against B- cell lymphomas [109]. Calgarotto and his research group revealed that a combination of polyphenols including green tea and quercetin reduced the tumor growth (HL-60) in human xenografted

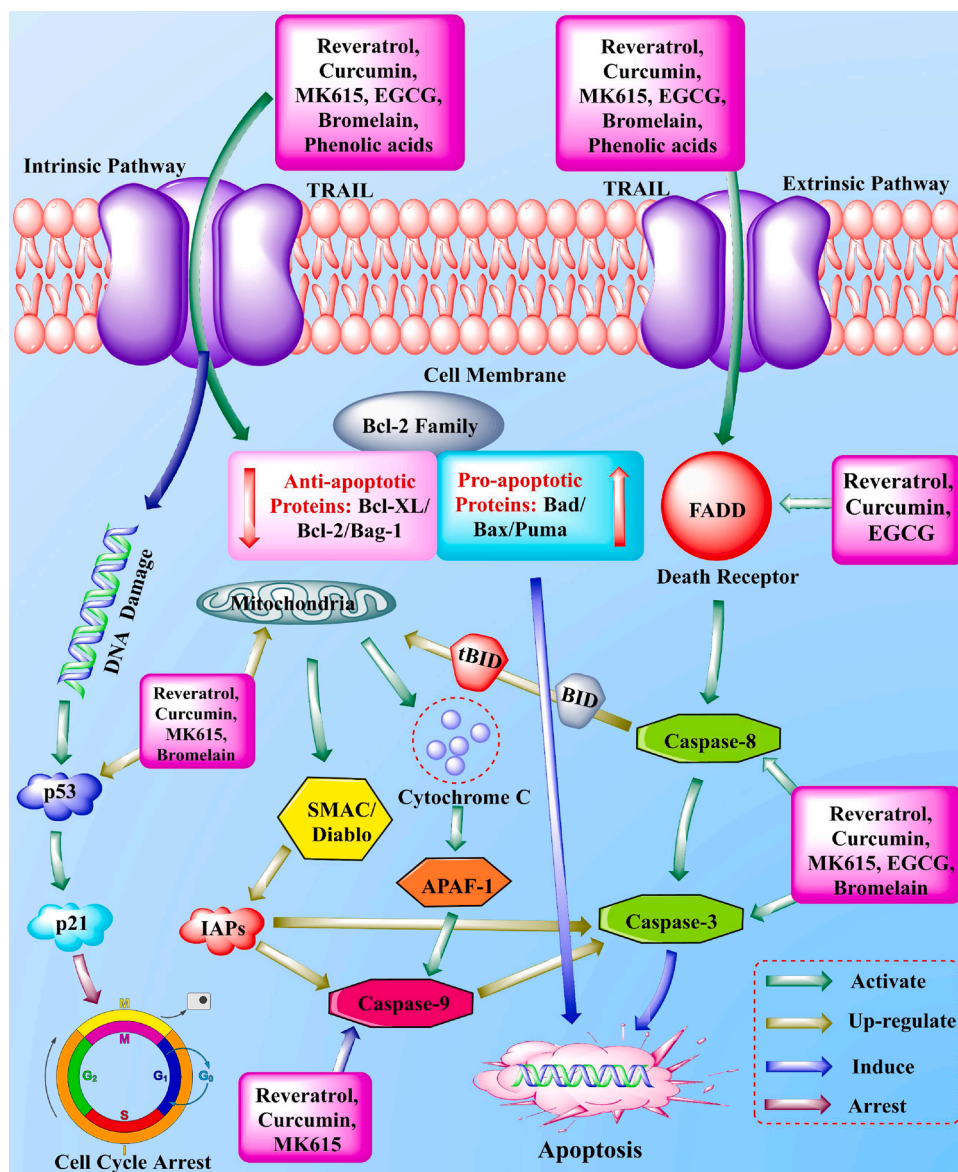


Fig. 2. The schematic diagram of apoptosis and cell cycle arrest by biomolecules in the treatment of lymphoma. TRAIL (Tumor-necrosis-factor Related Apoptosis-Inducing Ligand; purple); Bax (Bcl-2-associated X protein; sky box); Bcl-2 (B-cell lymphoma 2; pink box); Bak (Bcl-2 homologous antagonist killer; sky box); FADD (Fas-Associated protein with Death Domains; red circle); SMAC (Second Mitochondrial Activator of Caspase; yellow); BID [BH3 (Bcl-2 Homology-3)- Interacting Domain death agonist; light blue]; and APAF-1 (Apoptosis Activator Factor-1; orange); and IAPs (Inhibitor of Apoptosis proteins; red box).

SCID mice *via* inhibiting the activities of anti-apoptotic proteins (Bcl-2 and Mcl-1) and increase the expression of pro-apoptotic proteins including Bax. They also reported that consumption of green tea and quercetin activates the caspase-3 pathway to a greater extent and arrests the cell cycle (G1 phase) in HL-60 xenografted cells [29]. In another study, it has been proven that consumption of epigallocatechin-3-gallate (EGCG) showed significant effects against aggressive B-cell lymphoma by inhibiting the cell-proliferation and inducing apoptosis [110]. Similarly, Mirtavoos-Mahyari et al. [111] found that higher consumption of green tea (nonfermented) was associated with a 39 % reduced risk of NHL.

The summary of different plants with their active compounds against different lymphomas are presented in Table 3, and the schematic representation of apoptosis and cell cycle arrest in lymphomas are depicted in Fig. 2.

6. Conclusion

Lymphoma is considered a heterogeneous group of malignancies. Several conditions are exhibited by patients suffering from lymphoma, starting from mild fever to severe condition. However, timely diagnosis

is required to subclassify and proper treatment plans against lymphoma. Biochemical parameters (serum LDH and cytokine estimation) and biopsy techniques are used for diagnostic purposes. Genetic disordered, bacterial, and viral infections and environmental factors are associated with the rising incidences of lymphoma. According to the data of present review, it is revealed that natural products (*Curcuma longa*, *Prunus mume*, *Vitis vinifera*, *Annona reticulata*, *Ananas comosus*, and *Camellia sinensis*) proved as a valuable therapeutic option against different types of lymphomas including NHL, HL, B-cell lymphoma, Dalton’s lymphoma, and Burkitt lymphoma by inhibiting cell-proliferation, metastasis, inducing apoptosis, angiogenesis, suppressing PI3K/Akt/mTOR pathway, and activation of NF-κB and STAT3 pathways. The *in-vitro* study of *Annona* fruit (*Annona muricata*) showed the highest results of about 95 % in the reduction of lymphoma cell line Romas-1 in a dose-dependent manner. Similarly, *in vivo* study of curcumin reported a 50.5 % of reduction of Hodgkin lymphoma by inhibiting cell-proliferation proteins (Mcl-1 and XIAP). The present review findings proved natural products as a promising option to treat lymphoma and lead to the development of other natural active drugs. However, further examination is required to conduct the clinical trials to understand the exact mechanism of action of natural drugs on human body and to

isolate the more active constituents from other medicinal plants. It is also recommended to conduct further study on other species of plants in the near future to find optional human benefits concerning lymphomas.

Consent for publication

For this type of study informed consent is not required.

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CRedit authorship contribution statement

A. Hazafa, Hafiz M.N. Iqbal and S. Ahmad: Conceived the presented data, Writing - original draft, Software, Supervision. **A. Batool, M. Imran and H.A. Khan:** Developed the theory, Formal analysis, & Investigation. **Hafiz M.Z. Abideen and A. Zafar:** Revision and Editing.

Declaration of Competing Interest

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References

- [1] R. Nair, N. Arora, M.K. Mallath, Epidemiology of non-Hodgkin's lymphoma in India, *Oncology* 91 (Suppl. 1) (2016) 18–25.
- [2] J.L. Biecler, T.C. El-Galaly, M. Bøgsted, J. Jørgensen, P. de Nully Brown, C. B. Poulsen, J. Starklint, M.B. Juul, J.H. Christensen, P. Josefsson, Clinical prognostic scores are poor predictors of overall survival in various types of malignant lymphomas, *Leuk. Lymphoma* 60 (6) (2019) 1580–1583.
- [3] H. AlShaya, H. Li, A. Hamama, R.A. Siddiqui, Polyphenolic content and anti-oxidation activity in Annona fruit and its effects on lymphoma Cancer cells proliferation, *FASEB J.* 33 (1 supplement) (2019) 4711–4719.
- [4] A.C. Wotherspoon, M.-Q. Du, J. Spencer, Gastrointestinal Lymphoma, *Mucosal Immunology*, Elsevier, 2015, pp. 1737–1748.
- [5] J.O. Armitage, R.D. Gascoyne, M.A. Lunning, F. Cavalli, Non-hodgkin lymphoma, *Lancet* 390 (10091) (2017) 298–310.
- [6] S. Su, R. Liu, Y. Hu, G. Gong, W. Zhu, K. Wang, Neck lumps with enlarged lymph nodes: papillary thyroid Cancer and hodgkin lymphoma, *Ann. Case Rep. Images* 10 (2019) 2574–7754.
- [7] M. Dehghani, S. Haddadi, R. Vojdani, Signs, symptoms and complications of non-Hodgkin's lymphoma according to grade and stage in South Iran, *Asian Pac. J. Cancer Prev.* 16 (8) (2015) 3551–3557.
- [8] M. Intra, D.M. Fanianos, Breast Cancer in Lymphoma Survivors, *Breast Cancer*, Springer, 2017, pp. 399–414.
- [9] A. Miranda-Filho, M. Piñeros, A. Znaor, R. Marcos-Gragera, E. Steliarova-Foucher, F. Bray, Global patterns and trends in the incidence of non-Hodgkin lymphoma, *Cancer Causes Control* 30 (5) (2019) 489–499.
- [10] F. Ahmadi, A. Abadi, A. Baghestani, Trends of Non-Hodgkin Lymphoma cancer death rates with adjusting the effect of the human development index: the global assessment in 1990–2015, *World Cancer Res. J.* 6 (2019) e1325.
- [11] J. Ferlay, I. Soerjomataram, R. Dikshit, S. Eser, C. Mathers, M. Rebelo, D. M. Parkin, D. Forman, F. Bray, Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012, *Int. J. Cancer* 136 (5) (2015) E359–E386.
- [12] B. Togo, P. Togo, O. Koné, F. Traore, A. Doumbia, A. Touré, A. Diakité, O. Coulibaly, H. Diall, B. Maïga, Non hodgkin lymphomas (NHL) in the pediatric oncology unit of the gabriel touré teaching hospital, Bamako Mali, *Open J. Pediatr.* 9 (04) (2019) 309–317.
- [13] R.L. Siegel, K.D. Miller, A. Jemal, Cancer statistics, 2016, *CA-Cancer J. Clin.* 66 (1) (2016) 7–30.
- [14] H.M. Al-Shaya, H. Li, O.U. Beg, A.A. Hamama, S.M. Witiak, P. Kaseloo, R. A. Siddiqui, Phytochemical profile and antioxidant activity of annona fruit and its effect on lymphoma cell proliferation, *Food Sci. Nutr.* 8 (1) (2019) 58–68.
- [15] G.S. Calip, P.R. Patel, S. Adimadhyam, S. Xing, Z. Wu, K. Sweiss, G.T. Schumock, T.A. Lee, B.C.H. Chiu, Tumor necrosis factor-alpha inhibitors and risk of non-Hodgkin lymphoma in a cohort of adults with rheumatologic conditions, *Int. J. Cancer* 143 (5) (2018) 1062–1071.
- [16] A. Younes, *Handbook of Lymphoma*, Springer, 2016.
- [17] M.R. Drylewicz, M.P. Watkins, A.S. Shetty, M.F. Lin, A. Salter, N.L. Bartlett, W. D. Middleton, M. Yano, Formulating a treatment plan in suspected lymphoma: ultrasound-guided core needle biopsy versus core needle biopsy and fine-needle aspiration of peripheral lymph nodes, *J. Ultra. Med.* 38 (3) (2019) 581–586.
- [18] A. Johl, E. Lengfelder, W. Hiddemann, W. Klapper, G.L.-g.L.S. Group, Core needle biopsies and surgical excision biopsies in the diagnosis of lymphoma—experience at the Lymph Node Registry Kiel, *Ann. Hematol.* 95 (8) (2016) 1281–1286.
- [19] L.I. Levin, E.C. Breen, B.M. Birmann, J.L. Batista, L.I. Magpantay, Y. Li, R. F. Ambinder, N.E. Mueller, O. Martínez-Maza, Elevated serum levels of sCD30 and IL6 and detectable IL10 precede classical Hodgkin lymphoma diagnosis, *Cancer Epidemiol. Prev. Biom.* 26 (7) (2017) 1114–1123.
- [20] C.S. Chuen, N.M. Lazim, L.E. Haw, N.A.C. Jalil, H. Hazmi, Approach for paediatric cervical lymphadenopathy: tuberculosis versus lymphoma, *Bangladesh J. Med. Sci.* 18 (1) (2019) 149–152.
- [21] D.C. Hodgson, Long-term toxicity of chemotherapy and radiotherapy in lymphoma survivors: optimizing treatment for individual patients, *Clin. Adv. Hematol. Oncol.* 13 (2) (2015) 103–112.
- [22] C. Ajila, S. Brar, Role of Dietary Antioxidants in Cancer, *Nutrition, Diet and Cancer*, Springer, 2012, pp. 377–412.
- [23] F. Jubeen, A. Liaqat, F. Amjad, M. Sultan, S.Z. Iqbal, I. Sajid, M.B. Khan Niazi, F. Sher, Synthesis of 5-fluorouracil cocrystals with novel organic acids as cofomers and anticancer evaluation against HCT-116 colorectal cell lines, *Cryst. Growth Des.* 20 (4) (2020) 2406–2414.
- [24] F. Jubeen, A. Liaqat, M. Sultan, S.Z. Iqbal, I. Sajid, F. Sher, Green synthesis and biological evaluation of novel 5-fluorouracil derivatives as potent anticancer agents, *J. Saudi Pharm. Soc.* 27 (8) (2019) 1164–1173.
- [25] R. Debnath, D. Majumder, A. Singha, D. Ghosh, D. Maiti, Bromelain plus peroxidase from pineapple induces apoptosis via mitochondrial dependent pathway in lymphoma cells, *Int. J. Pharm. Sci. Res.* 9 (2018) 4610–4618.
- [26] S. Kim, S.S.A. An, Role of p53 isoforms and aggregations in cancer, *Medicine* 95 (26) (2016).
- [27] A. Oyenih, C. Smith, Are polyphenol antioxidants at the root of medicinal plant anti-cancer success? *J. Ethnopharmacol.* 229 (2019) 54–72.
- [28] A. Amini, S. Masoumi-Moghaddam, A. Ehteda, D.L. Morris, Bromelain and N-acetylcysteine inhibit proliferation and survival of gastrointestinal cancer cells *in vitro*: significance of combination therapy, *J. Exp. Clin. Cancer Res.* 33 (1) (2014) 92.
- [29] A.K. Calgarotto, V. Maso, G.C.F. Junior, A.E. Nowill, P. Latuf Filho, J. Vassallo, S. T.O. Saad, Antitumor activities of Quercetin and Green Tea in xenografts of human leukemia HL60 cells, *Sci. Rep.* 8 (1) (2018) 1–7.
- [30] M.K. Shanmugam, G. Rane, M.M. Kanchi, F. Arfuso, A. Chinnathambi, M. Zayed, S.A. Alharbi, B.K. Tan, A.P. Kumar, G. Sethi, The multifaceted role of curcumin in cancer prevention and treatment, *Molecules* 20 (2) (2015) 2728–2769.
- [31] A.R. Hussain, S. Uddin, R. Bu, O.S. Khan, S.O. Ahmed, M. Ahmed, K.S. Al-Kuraya, Resveratrol suppresses constitutive activation of AKT via generation of ROS and induces apoptosis in diffuse large B cell lymphoma cell lines, *PLoS One* 6 (9) (2011), e24703.
- [32] W.D. Lewis, S. Lilly, K.L. Jones, Lymphoma: diagnosis and treatment, *Am. Fam. Physician* 101 (1) (2020) 34–41.
- [33] H.W. Wang, J.P. Balakrishna, S. Pittaluga, E.S. Jaffe, Diagnosis of Hodgkin lymphoma in the modern era, *British J. Haematol.* 184 (1) (2019) 45–59.
- [34] R.N. Miranda, J.D. Khoury, L.J. Medeiros, Nodular Sclerosis Hodgkin Lymphoma, *Atlas of Lymph Node Pathology*, Springer, 2013, pp. 363–368.
- [35] R.N. Miranda, J.D. Khoury, L.J. Medeiros, Mixed Cellularity Hodgkin Lymphoma, *Atlas of Lymph Node Pathology*, Springer, 2013, pp. 373–376.
- [36] R.N. Miranda, J.D. Khoury, L.J. Medeiros, Lymphocyte-Depleted Hodgkin Lymphoma, *Atlas of Lymph Node Pathology*, Springer, 2013, pp. 377–380.
- [37] S.M. Ansell, Hodgkin lymphoma: 2018 update on diagnosis, risk-stratification, and management, *Am. J. Hematol.* 93 (5) (2018) 704–715.
- [38] Y.L. Boo, Y. Ting, H. Siew, D.F.S. Yap, S.G. Toh, S.M. Lim, Clinical features and treatment outcomes of Hodgkin lymphoma: a retrospective review in a Malaysian tertiary hospital, *Blood Res.* 54 (3) (2019) 210–217.
- [39] V.A. Morrison, Diffuse large B-Cell lymphomas in older adults, *Geriatr. Oncol.* 20 (2020) 583–612.
- [40] D. Tan, S.J. Horning, Follicular lymphoma: clinical features and treatment, *Hematol. Oncol. Clin. North Am.* 22 (5) (2008) 863–882.
- [41] R.S. Kelly, H. Kiviranta, I.A. Bergdahl, D. Palli, A.-S. Johansson, M. Botsivali, P. Vineis, R. Vermeulen, S.A. Kyrtopoulos, M. Chadeau-Hyam, Prediagnostic plasma concentrations of organochlorines and risk of B-cell non-Hodgkin lymphoma in environomarkers: a nested case-control study, *Environ. Health* 16 (1) (2017) 9.
- [42] M.E. Kadin, What cytokines can tell us about the pathogenesis of breast implant-associated anaplastic large cell lymphoma (BIA-ALCL), *Aesthetic Sur. J.* 39 (Supplement 1) (2019) S28–S35.
- [43] S.N. Chaudhry, A. Hazafa, M. Mumtaz, U. Kalsoom, S. Abbas, A. Kainaat, S. Bilal, N. Zafar, A. Siddique, A. Zafar, New insight on possible vaccine development against SARS-CoV-2, *Life Sci.* (2020), 118421.
- [44] S.M. Al-Khatib, N. Abdo, L.N. AL-Eitan, A.-H. Al-Mistarehi, D.J. Zahran, T. Z. Kewan, The Impact of IL-6 and IL-10 Gene Polymorphisms in Diffuse Large B-Cell Lymphoma Risk and Overall Survival in an Arab Population: A Case-Control Study, *Cancers* 12 (2) (2020) 382.
- [45] A. Miyajima, T. Kitamura, N. Harada, T. Yokota, K.-i. Arai, Cytokine receptors and signal transduction, *Ann. Rev. Immunol.* 10 (1) (1992) 295–331.
- [46] A. Hazafa, K. Ur-Rahman, I.-u.- Haq, N. Jahan, M. Mumtaz, M. Farman, H. Naeem, F. Abbas, M. Naeem, S. Sadiqa, The broad-spectrum antiviral recommendations for drug discovery against COVID-19, *Drug Metab. Rev.* 52 (3) (2020) 408–424.

- [47] F. Larousserie, D. Kebe, T. Huynh, A. Audebourg, J. Tamburini, B. Terris, O. Devergne, Evidence for IL-35 expression in diffuse large B-Cell lymphoma and impact on the patient's prognosis, *Front. Oncol.* 9 (2019) 563.
- [48] J.J. O'Shea, M. Gadina, R.M. Siegel, Cytokines and Cytokine Receptors, *Clinical Immunology*, Elsevier, 2019, pp. 127–155, e1.
- [49] A. Hazafa, A. Batool, S. Ahmad, M. Amjad, S.N. Chaudhry, J. Asad, H.F. Ghuman, H.M. Khan, M. Naeem, U. Ghani, Humanin: A mitochondrial-derived peptide in the treatment of apoptosis-related diseases, *Life Sci.* (2020), 118679.
- [50] A. Hazafa, M. Mumtaz, M.F. Farooq, S. Bilal, S.N. Chaudhry, M. Firdous, H. Naeem, M.O. Ullah, M. Yameen, M.S. Mukhtiar, CRISPR/Cas9: A powerful genome editing technique for the treatment of cancer cells with present challenges and future directions, *Life Sci.* (2020), 118525.
- [51] J. Ginn, J. Sacco, Y. Wong, A. Motsinger-Reif, R. Chun, L. Trepanier, Positive association between a glutathione-S-transferase polymorphism and lymphoma in dogs, *Vet. Comp. Oncol.* 12 (3) (2014) 227–236.
- [52] S. Nissar, A. Sameer, R. Rasool, N. Chowdhri, F. Rashid, Glutathione S transferases: biochemistry, polymorphism and role in colorectal carcinogenesis, *J. Carcinog. Mutagen.* 8 (2) (2017) 287–294.
- [53] L. Qin, H.-Y. Deng, S.-J. Chen, W. Wei, A meta-analysis on the relationship between hair dye and the incidence of non-hodgkin's lymphoma, *Med. Princ. Pract.* 28 (3) (2019) 222–230.
- [54] S. Yadlapati, P. Eftimiou, Autoimmune/inflammatory arthritis associated lymphomas: who is at risk? *Biomed Res. Int.* 2016 (2016), 8631061.
- [55] S. Hu, D. Zhou, Y. Wu, Y. Zhao, S. Wang, B. Han, M. Duan, J. Li, T. Zhu, J. Zhuang, Autoimmune disease-associated non-Hodgkin's lymphoma—a large retrospective study from China, *Ann. Hematol.* 98 (2) (2019) 445–455.
- [56] L.D. Kaulen, D. Galluzzo, P. Hui, F. Barbiero, P. Karschnia, A. Huttner, R. Fulbright, J.M. Baehring, Prognostic markers for immunodeficiency-associated primary central nervous system lymphoma, *J. Neurooncol.* 144 (1) (2019) 107–115.
- [57] Y.-f. Guo, J.-x. Pan, W.-h. Zhuang, Concurrent and reactivation of hepatitis B virus infection in diffuse large B-cell lymphoma: risk factors and survival outcome, *Infect. Agents Cancer* 13 (1) (2018) 40.
- [58] M. Hatakeyama, Malignant *Helicobacter Pylori*-associated Diseases: Gastric Cancer and MALT Lymphoma, *Helicobacter pylori in Human Diseases*, Springer, 2019, pp. 135–149.
- [59] V.J. Cogliano, R. Baan, K. Straif, Y. Grosse, B. Lauby-Secretan, F. El Ghissassi, V. Bouvard, L. Benbrahim-Tallaa, N. Guha, C. Freeman, Preventable exposures associated with human cancers, *J. Natl. Cancer Inst.* 103 (24) (2011) 1827–1839.
- [60] H. Hjalgrim, E. Engels, Infectious aetiology of Hodgkin and non-Hodgkin lymphomas: a review of the epidemiological evidence, *J. Int. Med.* 264 (6) (2008) 537–548.
- [61] A.M. Elasbali, H.H. Alharbi, Z. Al-Onzi, A. Hamza, E. Khalafalla, H.G. Ahmed, Epidemiology and patterns of malignant lymphoma in Northern Saudi Arabia, *Open J. Blood Dis.* 8 (04) (2018) 83–87.
- [62] H. Emami, S.A. Zadeegan, H.N. Bonaki, R. Erfanian, A case report of diffuse large B-cell lymphoma masquerading as necrotizing lymph nodes: the role of core needle biopsy for early diagnosis, *Int. J. Cancer Manag.* 11 (2) (2018) e9571.
- [63] A. Wilczynski, C. Görg, N. Timmesfeld, A. Ramaswamy, A. Neubauer, A. Burchert, C. Trenker, Value and diagnostic accuracy of ultrasound-guided full core needle biopsy in the diagnosis of lymphadenopathy: a retrospective evaluation of 793 cases, *J. Ultra. Med.* 39 (3) (2020) 559–567.
- [64] F. Knörr, C. Damm-Welk, S. Ruf, V.K. Singh, M. Zimmermann, A. Reiter, W. Woessmann, Blood cytokine concentrations in pediatric patients with anaplastic lymphoma kinase-positive anaplastic large cell lymphoma, *Haematologica* 103 (3) (2018) 477–485.
- [65] K. Augoff, A. Hryniewicz-Jankowska, R. Tabola, Lactate dehydrogenase 5: an old friend and a new hope in the war on cancer, *Cancer Lett.* 358 (1) (2015) 1–7.
- [66] E. Saba, Y.Y. Lee, M. Kim, H.-K. Kim, M.H. Rhee, Comparison of the antioxidant activities of various processed fruits and vegetables in APAP-induced oxidative stress in BALB/c mice, *대한의생명과학회지* 25 (3) (2019) 211–217.
- [67] D. Sinha, Role of different polyphenols in the treatment of cancer disease, *Asian J. Pharm. Pharmacol.* 6 (1) (2020) 1–19.
- [68] A. Hazafa, K.-U.- Rehman, N. Jahan, Z. Jabeen, The role of polyphenol (flavonoids) compounds in the treatment of cancer cells, *Nutr. Cancer* 72 (3) (2020) 386–397.
- [69] D. Angelino, J. Godos, F. Ghelfi, M. Tieri, L. Titta, A. Lafranconi, S. Marventano, E. Alonzo, A. Gambera, S. Sciacca, Fruit and vegetable consumption and health outcomes: an umbrella review of observational studies, *Int. J. Food Sci. Nutr.* 70 (6) (2019) 652–667.
- [70] P. Maiti, A. Plemmons, G.L. Dunbar, Combination treatment of berberine and solid lipid curcumin particles increased cell death and inhibited PI3K/Akt/mTOR pathway of human cultured glioblastoma cells more effectively than did individual treatments, *PLoS One* 14 (12) (2019).
- [71] Y. Li, A. Domina, G. Lim, T. Chang, T. Zhang, Evaluation of curcumin, a natural product in turmeric, on Burkitt lymphoma and acute myeloid leukemia cancer stem cell markers, *Future Oncol.* 14 (23) (2018) 2353–2360.
- [72] G. Mackenzie, N. Queisser, M.L. Wolfson, C.G. Fraga, A.M. Adamo, O. P, Curcumin induces cell-arrest and apoptosis in association with the inhibition of constitutively active NF-kappaB and STAT3 pathways in Hodgkin's lymphoma cells, *Int. J. Cancer* 123 (2008) 56–65.
- [73] J. Guorgui, R. Wang, G. Mattheolabakis, G.G. Mackenzie, Curcumin formulated in solid lipid nanoparticles has enhanced efficacy in Hodgkin's lymphoma in mice, *Arch. Biochem. Biophys.* 648 (2018) 12–19.
- [74] W. Guo, Y. Song, W. Song, Y. Liu, Z. Liu, D. Zhang, Z. Tang, O. Bai, Co-delivery of Doxorubicin and curcumin with polypeptide nanocarrier for Synergistic Lymphoma therapy, *Sci. Rep.* 10 (1) (2020) 1–16.
- [75] L. Chen, C.-Z. Zhan, T. Wang, H. You, R. Yao, Curcumin inhibits the proliferation, migration, invasion, and apoptosis of diffuse large B-Cell lymphoma cell line by regulating MiR-21/VHL Axis, *Yonsei Med. J.* 61 (1) (2020) 20–29.
- [76] L. Das, M. Vinayak, Curcumin modulates glycolytic metabolism and inflammatory cytokines via Nrf 2 in Dalton's lymphoma ascites cells *in vivo*, *Anti-Cancer Agent. Med. Chem.* 18 (12) (2018) 1779–1791.
- [77] S. Shishodia, H.M. Amin, R. Lai, B.B. Aggarwal, Curcumin (diferuloylmethane) inhibits constitutive NF-κB activation, induces G1/S arrest, suppresses proliferation, and induces apoptosis in mantle cell lymphoma, *Biochem. Pharmacol.* 70 (5) (2005) 700–713.
- [78] C. Park, C.-Y. Jin, G.-Y. Kim, Y.K. Jeong, W.-J. Kim, Y.H. Choi, Induction of apoptosis by ethanol extract of *Prunus mume* in U937 human leukemia cells through activation of caspases, *Oncol. Rep.* 26 (4) (2011) 987–993.
- [79] M. Inoue, Y. Honma, T. Urano, J. Suzumiyama, Japanese apricot extract (MK615) potentiates bendamustine-induced apoptosis via impairment of the DNA damage response in lymphoma cells, *Oncol. Lett.* 14 (1) (2017) 792–800.
- [80] C. Bailly, Anticancer properties of *Prunus mume* extracts (Chinese plum, Japanese apricot), *J. Ethnopharmacol.* 246 (2019), 112215.
- [81] R. Tsuji, H. Koizumi, D. Fujiwara, Effects of a plum (*Prunus mume* Siebold and Zucc.) ethanol extract on the immune system *in vivo* and *in vitro*, *Biosci. Biotechnol. Biochem.* 75 (10) (2011) 2011–2013.
- [82] M. Yanaki, M. Kobayashi, A. Aruga, M. Nomura, M. Ozaki, *In vivo* antitumor effects of MK615 led by PD-L1 downregulation, *Integr. Cancer Ther.* 17 (3) (2018) 646–653.
- [83] W.S. Al-Jahdari, H. Sakurai, Y. Yoshida, A. Mobaraki, Y. Suzuki, T. Nakano, MK615, a prospective anti-proliferative agent, enhances CD4/CD8 ratio after exposure to irradiation, *Int. J. Radiat. Biol.* 87 (1) (2011) 81–90.
- [84] R. Frazzi, M. Guardi, Cellular and molecular targets of resveratrol on lymphoma and leukemia cells, *Molecules* 22 (6) (2017) 885.
- [85] O.S. Khan, A.A. Bhat, R. Krishnankutty, R.M. Mohammad, S. Uddin, Therapeutic potential of resveratrol in lymphoid malignancies, *Nutr. Cancer* 68 (3) (2016) 365–373.
- [86] J. Meng, G. Liu, J. Song, L. Chen, A. Wang, X. Gao, Z. Wang, Preliminary results indicate resveratrol affects proliferation and apoptosis of leukemia cells by regulating PTEN/PI3K/AKT pathway, *Euro. rev. Med. Pharmacol. Sci.* 23 (2019) 4285–4292.
- [87] Y. Kong, G. Chen, Z. Xu, G. Yang, B. Li, X. Wu, W. Xiao, B. Xie, L. Hu, X. Sun, Pterostilbene induces apoptosis and cell cycle arrest in diffuse large B-cell lymphoma cells, *Sci. Rep.* 6 (2016) 37417.
- [88] S.J. Zunino, D.H. Storms, Resveratrol alters proliferative responses and apoptosis in human activated B lymphocytes *in vitro*, *J. Nutr.* 139 (8) (2009) 1603–1608.
- [89] F.F. Radwan, L. Zhang, A. Hossain, B.P. Doonan, J.M. God, A. Haque, Mechanisms regulating enhanced human leukocyte antigen class II-mediated CD4+ T cell recognition of human B-cell lymphoma by resveratrol, *Leuk. Lymphoma* 53 (2) (2012) 305–314.
- [90] Y.-C. Ko, C.-L. Chang, H.-F. Chien, C.-H. Wu, L.-I. Lin, Resveratrol enhances the expression of death receptor Fas/CD95 and induces differentiation and apoptosis in anaplastic large-cell lymphoma cells, *Cancer Lett.* 309 (1) (2011) 46–53.
- [91] A.C. Faber, T.C. Chiles, Resveratrol induces apoptosis in transformed follicular lymphoma OCI-LY8 cells: evidence for a novel mechanism involving inhibition of BCL6 signaling, *Int. J. Oncol.* 29 (6) (2006) 1561–1566.
- [92] P. Jara, J. Spies, C. Cárcamo, Y. Arancibia, G. Vargas, C. Martin, M. Salas, C. Otth, A. Zambrano, The effect of resveratrol on cell viability in the Burkitt's lymphoma cell line Ramos, *Molecules* 23 (1) (2018) 14.
- [93] R. Pavan, S. Jain, A. Kumar, Properties and therapeutic application of bromelain: a review, *Biotechnol. Res. Int.* 2012 (2012), 976203.
- [94] P. Gross, H. Seelert, P. Meiser, R. Müller, Characterization of bromelain indicates a molar excess of inhibitor vs enzyme molecules, a Jacalin-like lectin and Maillard reaction products, *J. Pharm. Biomed. Anal.* 181 (2020), 113075.
- [95] E. Misran, A. Idris, S.H.M. Sarip, H. Ya'akob, Properties of bromelain extract from different parts of the pineapple variety Morris, *Biocatal. Agric. Biotechnol.* 18 (2019), 101095.
- [96] A. Amini, S. Masoumi-Moghaddam, D.L. Morris, Bromelain, Utility of Bromelain and N-Acetylcysteine in Treatment of Peritoneal Dissemination of Gastrointestinal Mucin-Producing Malignancies, *Springer*, 2016, pp. 63–80.
- [97] E.R. Secor Jr, A. Singh, L.A. Guernsey, J.T. McNamara, L. Zhan, N. Maulik, R. S. Thrall, Bromelain treatment reduces CD25 expression on activated CD4+ T cells *in vitro*, *Int. Immunopharmacol.* 9 (3) (2009) 340–346.
- [98] S.-i. Kurata, Selective activation of p38 MAPK cascade and mitotic arrest caused by low level oxidative stress, *J. Biol. Chem.* 275 (31) (2000) 23413–23416.
- [99] R. Debnath, D. Majumder, P. Nath, D. Ghosh, D. Maiti, Bromelain plus peroxidase reduces non-Hodgkin lymphoma progression *in vivo* via up-regulation of antioxidant enzymes and modulating apoptotic protein expression, *Nutr. Cancer* 72 (7) (2020) 1200–1210.
- [100] A.I. Yajid, H.S. Ab Rahman, M.P.K. Wong, W.Z.W. Zain, Potential benefits of *Annona muricata* in combating cancer: a review, *Malays. J. Med. Sci.* 25 (1) (2018) 5–15.
- [101] A.V. Coria-Téllez, E. Montalvo-González, E.M. Yahia, E.N. Obledo-Vázquez, *Annona muricata*: a comprehensive review on its traditional medicinal uses, phytochemicals, pharmacological activities, mechanisms of action and toxicity, *Arabian J. Chem.* 11 (5) (2018) 662–691.
- [102] D. Nakano, K. Ishitsuka, M. Kamikawa, M. Matsuda, R. Tsuchihashi, M. Okawa, H. Okabe, K. Tamura, J. Kinjo, Screening of promising chemotherapeutic

- candidates from plants against human adult T-cell leukemia/lymphoma (III), *J. Nat. Med.* 67 (4) (2013) 894–903.
- [103] P. Nalini, B. Durairaj, Antitumor potential of hydroethanolic extract of *Annona muricata* leaves against Dalton's lymphoma ASCITES-INDUCED tumor in mice, *Asian J. Pharm. Clin. Res.* 11 (3) (2018) 364–367.
- [104] F. Calzada, J. Ramirez-Santos, M. Valdes, N. Garcia-Hernandez, E. Pina-Jiménez, R.M. Ordoñez-Razo, Evaluation of acute oral toxicity, brine shrimp lethality, and antilymphoma activity of Geranylgeraniol and *Annona macroprophyllata* leaf extracts, *Rev. Bras. Farmacogn.* 30 (2020) 301–304.
- [105] B. Vikas, S. Anil, P. Remani, Cytotoxicity profiling of *Annona squamosa* in Cancer cell lines, *Asian Pac. J. Cancer Prev.* 20 (9) (2019) 2831.
- [106] L. Liu, L. Hou, S. Gu, X. Zuo, D. Meng, M. Luo, X. Zhang, S. Huang, X. Zhao, Molecular mechanism of epigallocatechin-3-gallate in human esophageal squamous cell carcinoma *in vitro* and *in vivo*, *Oncol. Rep.* 33 (1) (2015) 297–303.
- [107] Y.H. Kim, Y.-S. Won, X. Yang, M. Kumazoe, S. Yamashita, A. Hara, A. Takagaki, K. Goto, F. Nanjo, H. Tachibana, Green tea catechin metabolites exert immunoregulatory effects on CD4+ T cell and natural killer cell activities, *J. Agri. Food Chem.* 64 (18) (2016) 3591–3597.
- [108] P. Xu, F. Yan, Y. Zhao, X. Chen, S. Sun, Y. Wang, L. Ying, Green tea polyphenol EGCG attenuates MDSCs-mediated immunosuppression through canonical and non-canonical pathways in a 4T1 murine breast Cancer model, *Nutrients* 12 (4) (2020) 1042.
- [109] D. Przystupski, M.J. Niemczura, A. Górská, S. Supplitt, K. Kotowski, P. Wawryka, P. Rozborska, K. Woźniak, O. Michel, A. Kielbik, In search of panacea—review of recent studies concerning nature-derived anticancer agents, *Nutrients* 11 (6) (2019) 1426.
- [110] S. Parodi, I. Santi, E. Marani, C. Casella, A. Puppo, E. Garrone, V. Fontana, E. Stagnaro, Lifestyle factors and risk of leukemia and non-Hodgkin's lymphoma: a case-control study, *Cancer Causes Control* 27 (3) (2016) 367–375.
- [111] H. Mirtavoos-Mahyari, P. Salehipour, M. Parohan, A. Sadeghi, Effects of coffee, black tea and green tea consumption on the risk of non-hodgkin's lymphoma: a systematic review and dose-response meta-analysis of observational studies, *Nutr. Cancer* 71 (6) (2019) 887–897.
- [112] Z. Batool, N. Ali, H. Amin, S. Basharat, S. Hamayun, M. Idrees, Clinicopathological features of Hodgkin Lymphoma in patients of Peshawar region, *KJMS* 12 (1) (2019) 12.
- [113] A. Wahed, A. Quesada, A. Dasgupta, Hematology and Coagulation: a Comprehensive Review for Board Preparation, Certification and Clinical Practice, Academic Press, 2019.
- [114] M. Al-Hamadani, T.M. Habermann, J.R. Cerhan, W.R. Macon, M.J. Maurer, R. S. Go, Non-H odgkin lymphoma subtype distribution, geodemographic patterns, and survival in the US: a longitudinal analysis of the National Cancer D ata B ase from 1998 to 2011, *Am. J. Hematol.* 90 (9) (2015) 790–795.
- [115] A.M. Perry, J. Diebold, B.N. Nathwani, K.A. MacLennan, H.K. Müller-Hermelink, M. Bast, E. Boilesen, J.O. Armitage, D.D. Weisenburger, Non-Hodgkin lymphoma in the developing world: review of 4539 cases from the International Non-Hodgkin Lymphoma Classification Project, *Haematologica* 101 (10) (2016) 1244–1250.
- [116] C. Page, E. Saunders, Z. Najim, O. Firth, H. Owens, N. Gill, A. Hodson, M. Prahladan, Mantle Cell Lymphoma (MCL): a single centre experience on the outcome of MCL before and after the emergence of novel THERAPIES.: PB2018, *HemaSphere* 3 (2019) 912–913.
- [117] J.H. Won, S.M. Kim, J.W. Kim, J.H. Park, J.Y. Kim, Clinical features, treatment and outcomes of colorectal mucosa-associated lymphoid tissue (MALT) lymphoma: literature reviews published in English between 1993 and 2017, *Cancer Manag. Res.* 11 (2019) 8577.
- [118] A.S. Freedman, J.C. Aster, Clinical Manifestations, Pathologic Features, and Diagnosis of Lymphoplasmacytic Lymphoma, U: UpToDate, Post TW ur. UpToDate [Internet], Waltham, MA, 2017. UpToDate.
- [119] H. Yang, Z. Du, W. Wang, M. Song, K. Sanidad, E. Sukamtoh, J. Zheng, L. Tian, H. Xiao, Z. Liu, Structure-activity relationship of curcumin: role of the methoxy group in anti-inflammatory and anticolic effects of curcumin, *J. Agric. Food Chem.* 65 (22) (2017) 4509–4515.
- [120] S. Sahu, R. Li, P.K. Kadeyala, S. Liu, M. Schachner, The human natural killer-1 (HNK-1) glycan mimetic ursolic acid promotes functional recovery after spinal cord injury in mouse, *J. Nutr. Biochem.* 55 (2018) 219–228.
- [121] A. Saneja, L. Sharma, R.D. Dubey, M.J. Minto, A. Singh, A. Kumar, P.L. Sangwan, S.A. Tasaduq, G. Singh, D.M. Mondhe, Synthesis, characterization and augmented anticancer potential of PEG-betulinic acid conjugate, *Mater. Sci. Eng. C.* 73 (2017) 616–626.
- [122] H.-H. Song, H.W. Ryu, H.-S. Kim, C.-S. Kim, H.-J. Hyun, H.-K. Lee, S.-R. Oh, A metabolomics approach to identify factors influencing their activity relative to oleonic acid contents in Korean mistletoe types, *J. Funct. Food.* 22 (2016) 64–72.
- [123] N.S. Alamolhodaei, A.M. Tsatsakis, M. Ramezani, A.W. Hayes, G. Karimi, Resveratrol as MDR reversion molecule in breast cancer: An overview, *Food Chem. Toxicol.* 103 (2017) 223–232.
- [124] X. Jun-hui, C. Hui-juan, Z. Bin, Y. Hui, The mechanistic effect of bromelain and papain on tenderization in jumbo squid (*Dosidicus gigas*) muscle, *Food Res. Int.* 131 (2020), 108991.
- [125] B. Liu, W. Yan, Lipophilization of EGCG and effects on antioxidant activities, *Food Chem.* 272 (2019) 663–669.
- [126] P. Meghana, J.R.S. Kumar, T. Sandeep, N. Prashanth, H. Kumaraswamy, Acetogenins as potential checkpoint-2 kinase inhibitors: an *in silico* analysis, *J. Proteins Proteom.* 10 (4) (2019) 299–311.
- [127] H. Kai, E. Akamatsu, E. Torii, H. Kodama, C. Yukizaki, Y. Sakakibara, M. Suiko, K. Morishita, H. Kataoka, K. Matsuno, Inhibition of proliferation by agricultural plant extracts in seven human adult T-cell leukaemia (ATL)-related cell lines, *J. Nat. Med.* 65 (3-4) (2011) 651–655.
- [128] R. Baez, M.T. Lopes, C.E. Salas, M. Hernandez, *In vivo* antitumoral activity of stem pineapple (*Ananas comosus*) bromelain, *Planta Med.* 73 (13) (2007) 1377–1383.