

<http://www.pjbs.org>

**PJBS**

ISSN 1028-8880

**Pakistan  
Journal of Biological Sciences**

**ANSI***net*

Asian Network for Scientific Information  
308 Lasani Town, Sargodha Road, Faisalabad - Pakistan



## Research Article

# Anti-proliferation and Apoptosis Induction of Aqueous Leaf Extract of *Carica papaya* L. on Human Breast Cancer Cells MCF-7

<sup>1</sup>Fatma Zuhrotun Nisa, <sup>2</sup>Mary Astuti, <sup>2</sup>Agnes Murdiati and <sup>1</sup>Sofia Mubarika Haryana

<sup>1</sup>Department of Health and Nutrition, Faculty of Medicine, Universitas Gadjah Mada, Jl Farmako Sekip Utara, 55281 Yogyakarta, Indonesia

<sup>2</sup>Faculty of Agriculture Technology, Universitas Gadjah Mada, Jl Flora No. 1, 55281 Yogyakarta, Indonesia

## Abstract

**Background and Objective:** Breast cancer is the most frequently diagnosed cancer in women. Chemotherapy is the main method of breast cancer treatment but there are side effects. *Carica papaya* leaves is vegetable foods consumed by most people of Indonesia have potential as anticancer. The aim of this study was to investigate anti-proliferative and apoptotic induced effect of aqueous papaya leaves extracts on human breast cancer cell lines MCF-7. **Materials and Methods:** Inhibitory on cell proliferation was measured by MTT assay while apoptosis induction was measured using Annexin V. **Results:** The results showed that papaya leaf can inhibit the proliferation of human breast cancer cells MCF-7 with  $IC_{50}$  in  $1319.25 \mu\text{g mL}^{-1}$ . The  $IC_{50}$  values of papaya leaf extract was higher than the  $IC_{50}$  value quercetin and doxorubicin. Papaya leaf extract can also induce apoptosis of breast cancer cells MCF-7 about 22.54% for concentration  $659.63 \mu\text{g mL}^{-1}$  and about 20.73% for concentration  $329.81 \mu\text{g mL}^{-1}$ . The percentage of cell apoptosis of papaya leaf extract lower than doxorubicin but higher than quercetin. **Conclusion:** This study indicated that papaya leaf extract have potential as anticancer through mechanism anti-proliferation and apoptosis induction.

**Key words:** *Carica papaya*, leaf, breast cancer, proliferation, apoptosis, extract, MTT assay, MCF-7 cell line

**Received:** September 10, 2016

**Accepted:** November 12, 2016

**Published:** December 15, 2016

**Citation:** Fatma Zuhrotun Nisa, Mary Astuti, Agnes Murdiati and Sofia Mubarika Haryana, 2017. Anti-proliferation and apoptosis induction of aqueous leaf extract of *Carica papaya* L. on human breast cancer cells MCF-7. Pak. J. Biol. Sci., 20: 36-41.

**Corresponding Author:** Fatma Zuhrotun Nisa, Department of Health and Nutrition, Faculty of Medicine, Universitas Gadjah Mada, Jl Farmako Sekip Utara, 55281 Yogyakarta, Indonesia

**Copyright:** © 2017 Fatma Zuhrotun Nisa *et al.* This is an open access article distributed under the terms of the creative commons attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

**Competing Interest:** The authors have declared that no competing interest exists.

**Data Availability:** All relevant data are within the paper and its supporting information files.

## INTRODUCTION

Cancer is a degenerative disease that is a public health problem in the world. Based on the results of RISKESDAS was known cancer incidence in Indonesia reached 1.4% per 1,000 respondents<sup>1</sup>. Breast cancer is the most frequently diagnosed cancer in women<sup>2</sup>. Chemotherapy is the main method of breast cancer treatment. However, since there are side effects of anticancer drugs<sup>3</sup>, natural products such as herbs have been used as an alternative therapy<sup>4</sup>. Moreover, consumption of phytoestrogens found in foods such as soy beans is associated with a lower risk of breast cancer<sup>5</sup>.

*Carica papaya* leaf has been used as a folk medicine for a variety of ailments such as healing of burns, relief of asthma symptoms, treatment of intestinal worms, treatment of digestion problems, fever control and treatment of amoebic dysentery<sup>6-8</sup>. Papaya leaf has also been used to increase appetite, ease menstrual pain and relieve nausea<sup>9</sup>. A recent study found that papaya leaf extract could prevent growth of cancer cells including breast cancer<sup>10</sup>. However, because the study investigated all cancer cells not specific cancer cell and the study only tested cytotoxic activity, this study will investigate specific breast cancer cell lines and also tested the apoptotic induced. The aim of this study was to test anti-proliferative and apoptotic induced effect of papaya leaves extracts on human breast cancer cell lines MCF-7.

## MATERIALS AND METHODS

**Material:** *Carica papaya* leaves are used in this study were obtained from local farmers in Yogyakarta with variety Grendel. Sampling of *Carica papaya* leaves in this study based on observation of the number of leaves in the tree then divided three parts. Number of papaya leaves in every tree ranged between 18-25 leaves. Papaya leaves sample taken from the 7th or 8th.

**Sample preparation:** Papaya leaves are dried in an oven of 60°C for 3 h 3 times then milled. About 1 g of dried papaya leaves milled put in 20 mL of solvent and stirred. Solvents used in this study are methanol, ethanol 70% and water. The extraction method used is the percentage watt microwave/heat by 50% by setting 4 sec on and 60 sec off for three times then performed filtering using Whatman No. 1 filter paper and then stored in a refrigerator for further testing<sup>11</sup>.

**Determination of cell proliferation:** Inhibitory of cell proliferation test was performed using

3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl-tetrazoliumbromide (MTT) assay according to Ebada *et al.*<sup>12</sup>. The MCF-7 cell (breast cancer cell) were cultured in DMEM medium, supplemented with 10% fetal bovine serum, 1% fungi zone and 2% penicillin-streptomycin. The cells were maintained at 37°C in a moisture saturated atmosphere containing 5% CO<sub>2</sub>. The cells were seeded at density of 2 × 10<sup>4</sup> cells per well in 100 µL of medium and allowed to attach overnight. After the cells were grown to about 80% confluence, treatments were initiated by supplementing serial dilution of 3000 µg mL<sup>-1</sup> for papaya leaves extract, 100 µg mL<sup>-1</sup> for quercetin and 10 µg mL<sup>-1</sup> for doxorubicin. All treatments were conducted in three replicates. The inhibition percentage of cells growth were calculated with formula: A-B/A × 100%, where A = Control cell absorbance, B = Compounds absorbance, the inhibition concentration 50 (IC<sub>50</sub>) value is defined as the concentrations of compound which inhibited 50% of the cell growth. The IC<sub>50</sub> value was determined by regression linear equation.

**Determination of apoptotic cell:** Determination of apoptotic cell was conducted using Annexin V staining kit according to Elmore<sup>13</sup>. Briefly, the MCF-7 cell was seeded at a final density of 5 × 10<sup>5</sup> cells per well in 3 wells micro culture and incubated for 12 h in CO<sub>2</sub> incubator (37°C, 5% of CO<sub>2</sub> flow). The extract were added to cells at 659.63 and 329.81 µg mL<sup>-1</sup> and incubated for 24 h. After 24 h of incubation, cells were harvested by the addition of trypsin, centrifuged for 5 min at 1000 rpm and finally washed with PBS. Finally, the cells were resuspended in 100 µL of Annexin-V-FLUOS staining kit (Roche) then incubated in dark room for 10 min at 20-25°C. Typical histogram of apoptotic cell was performed using flow cytometer. Doxorubicin was used as a positive control.

## RESULTS

The inhibition of proliferation of MCF-7 cells were tested by MTT assay *in vitro*. The principle of this method is reacting the bioactive compound with cancer cells. Conversion of tetrazolium salt (MTT) into formazan blue is found only in cells that are still alive and the amount of formazan produced is proportional to the number of existing living cells. Thus MTT assay was used to test potential anti-proliferative activity of the extract<sup>14</sup>. The percentage of death of human breast cancer cells MCF-7 with serial dilution of papaya leaf extract 3000 µg mL<sup>-1</sup>, quercetin 50 µg mL<sup>-1</sup> and doxorubicin 6.25 µg mL<sup>-1</sup> for 24 h incubation shown in Fig. 1-3.

This study shows the percentage of MCF-7 cell death is influenced by the concentration of the extract or samples. The greater concentration given on MCF-7 cells the number of

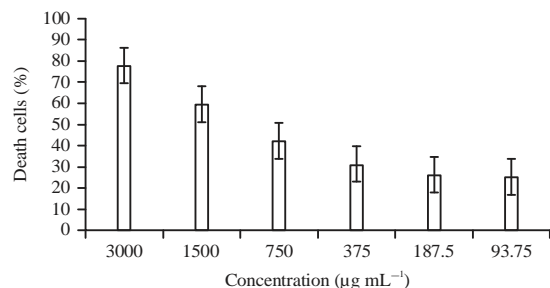


Fig. 1: Percentage death cells of MCF-7 by aqueous papaya leaf extract

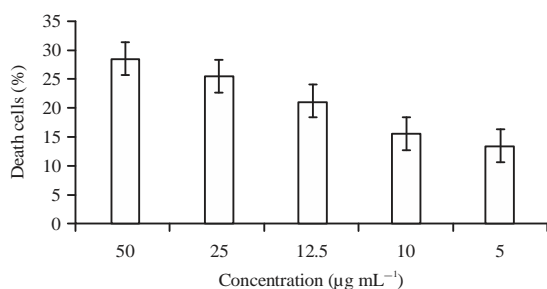


Fig. 2: Percentage death cells of MCF-7 by quercetin

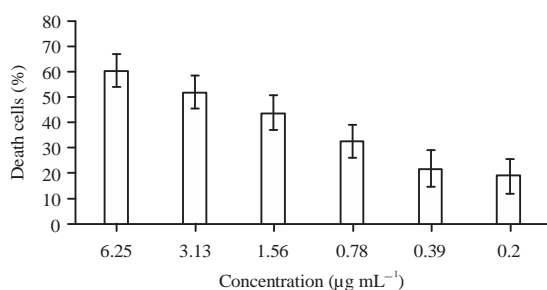


Fig. 3: Percentage death cells of MCF-7 by doxorubicin

Table 1: Inhibitory concentration 50% (IC<sub>50</sub>) value of extract, quercetin and doxorubicin

Samples	IC <sub>50</sub> (µg mL <sup>-1</sup> )
Extract	1319.25
Quercetin	111.36
Doxorubicin	3.87

dead cells increased. The results showed that the aqueous extract of papaya leaves has potential as an anti-proliferation in MCF-7 cells as evidenced by the increasing number of cell death compared to control.

Inhibition of aqueous papaya leaf extract on proliferation breast cancer cells MCF-7 was indicated by inhibitory concentration (IC<sub>50</sub>) value (Table 1). The IC<sub>50</sub> value were calculated by the linear regression equation from the data of Fig. 1-3. The linear regression equation obtained is:

Table 2: Percentage of apoptotic of human breast cancer cells MCF-7 by extract and doxorubicin

Samples	Dose (µg mL <sup>-1</sup> )	Apoptotic (%)
Extract	659.63	22.54±0.21
	329.81	20.73±0.06
Quercetin	55.68	3.48±0.18
	27.84	3.26±0.13
Doxo	1.94	86.80±0.22
	0.97	78.07±0.25

Data were expressed as Mean±SD, (n = 3)

$$y = 0.0186x + 25.462 \text{ with } r = 0.9676$$

The IC<sub>50</sub> value of aqueous extract of papaya leaf calculated using the equation was 1319.25 µg mL<sup>-1</sup>. The IC<sub>50</sub> of quercetin and doxorubicin were also measured for comparison as positive control. Quercetin is a flavonoid compounds contained in papaya shoots<sup>15</sup> while doxorubicin is a chemotherapeutic drug commonly given for breast cancer treatment. The IC<sub>50</sub> value of quercetin found 111.36 µg mL<sup>-1</sup> and IC<sub>50</sub> value of doxorubicin found 3.87 µg mL<sup>-1</sup>. The IC<sub>50</sub> values of papaya leaves extract is higher than quercetin and doxorubicin. It means that papaya leaf extract has anti-proliferative potency lower than quercetin and doxorubicin.

The induction of apoptosis in MCF-7 cells by the aqueous extract of *Carica papaya* L., leaves was monitored using flow cytometry. Results demonstrated the occurrence of cell death in MCF-7 cells following addition of the extract to the culture medium. Apoptosis was induced when cells were treated with 659.63 and 329.81 µg mL<sup>-1</sup> extract. The percentage of apoptotic of human breast cancer cells MCF-7 shown in Table 2.

*Carica papaya* leaves extract can induce apoptosis of MCF-7 cell line 22.54% for concentration 659.63 µg mL<sup>-1</sup> and 20.54% for concentration 329.81 µg mL<sup>-1</sup>. The percentage of apoptotic of MCF-7 by papaya leaves extract is lower than doxorubicin but is higher than quercetin.

## DISCUSSION

Potential of papaya leaves extract as antiproliferation of breast cancer cells is suspected due to the content of bioactive compounds flavonoids. Based on preliminary research it is known that papaya leaf extract containing flavonoids quercetin of 3.39 mg g<sup>-1</sup> and papaya leaf extract also has antioxidant activity. Antioxidants are known to reduce the risk of cancer in both research laboratory and epidemiological research through its ability to reduce damage caused by free radicals<sup>16</sup>. Supplements of vitamin C and E in breast cancer

shown to cause cell differentiation and apoptosis and inhibit tumor progression<sup>17-20</sup>. The results of epidemiological studies have shown that antioxidant supplements may reduce the risk of breast cancer recurrence or prevent the growth of breast cancer<sup>21</sup>. Rapid changes in diet and lifestyle, may influence heritability of the variant phenotypes that are dependent on the nutraceutical or functional food supplementation for their expression. It is possible to recognize the interaction of specific nutraceuticals, with the genetic code possessed by all nucleated cells<sup>22</sup>.

Mechanism of papaya leaf extract to inhibit the proliferation of breast cancer cells MCF-7 allegedly through the reduction of Reactive Oxygen Species (ROS) by antioxidant compounds contained in papaya leaf extract impact on the decrease of nuclear factor- $\kappa$ B (NF- $\kappa$ B) and impact on the gene expression associated with oxidative stress in MCF-7 cells such as COX-2, AP-1, Bcl-2 and Bcl-XL which can inhibit the proliferation of breast cancer cells MCF-7<sup>23-27</sup>. Decrease of NF- $\kappa$ B also resulted in decreased the expression of cyclin D1 and increased tumor suppressors such as p27, p21 and p53, causing cell cycle arrest which inhibit cancer cell proliferation<sup>28-33</sup>. The IC<sub>50</sub> value can be predicted by ligand-receptor binding interactions for protein kinase (CK2) employing quantitative structure active relationship (QSAR) model<sup>34</sup>.

Cells die by apoptosis when cells become old or damaged, necrosis or a combination of the two and are replaced with new cells. On the other hand, cancer cells are immortal since they are resistant to apoptosis. Chemotherapy kills cancer cells through apoptosis and/or necrosis<sup>35</sup>. Potential of aqueous extract of papaya leaves in triggering apoptosis probably caused by flavonoids. Flavonoids can stimulate apoptosis through multiple mechanisms include inhibition of the activity of DNA topoisomerase I/II, modulation of signaling pathways, decreasing gene expression of Bcl-2 and Bcl-XL, increasing the gene expression of Bax and Bak as well as the activation of endonucleases<sup>36</sup>. Apoptosis type 1 programmed cell death is a normal physiological process, however defects in apoptosis is a major cause of cancer<sup>13</sup>. The apoptotic mechanism is often used as a criterion for discovering new anticancer agents. Several natural compounds such as quercetin and curcumin have shown apoptotic-inducing properties<sup>37</sup>. Apoptosis is characterized by specific morphological and biochemical changes of cells, including cell shrinkage, nuclear condensation and DNA fragmentation, dynamic membrane blebbing and loss of cell adhesion, phosphatidylserine externalization and intracellular specific proteolysis<sup>38-41</sup>.

## CONCLUSION

In conclusion, our data indicated that the *Carica papaya* leaves extract specifically reduced viability of human breast cancer cells MCF-7. *Carica papaya* leaves extract have potential as anti-proliferative and apoptotic induction. However, the mechanism of the action is still unclear. Thus, further investigations including isolation of individual active flavonoid and elucidation of the molecular mechanisms involved are needed to fully understand the active ingredient and potential of *Carica papaya* leaves as a chemopreventive food.

## SIGNIFICANT STATEMENTS

Cancer is a degenerative disease that is a public health problem in the world. Breast cancer is the most frequently diagnosed cancer in women. Chemotherapy is the main method of breast cancer treatment but there are side effects. *Carica papaya* leaf is vegetable that usually consumed by Indonesian people. A recent study found that papaya leaf extract could prevent growth of cancer cells including breast cancer. Because the study investigated all cancer cells and only tested cytotoxic activity. This study will investigate specific breast cancer cell lines and also tested the apoptotic induced. This study can solve the problem for women to prevent breast cancer and for breast cancer patient to inhibit cancer cell proliferation by functional food.

## ACKNOWLEDGMENT

Researchers acknowledge the Health and Nutrition Department for their financial support in this research and LPPT Universitas Gadjah Mada for their facilities support.

## REFERENCES

1. KKRI., 2013. [Basic health research 2013]. Badan Penelitian Dan Pengembangan Kesehatan, Kementerian Kesehatan Republik Indonesia. <http://www.depkes.go.id/resources/download/general/Hasil%20Riskseddas%202013.pdf>, (In Indonesian)
2. Siegel, R., J. Ma, Z. Zou and A. Jemal, 2014. Cancer statistics, 2014. CA: Cancer J. Clin., 64: 9-29.
3. Abbasalipourkabir, R., A. Salehzadeh and R. Abdullah, 2010. Antitumor activity of tamoxifen loaded solid lipid nanoparticles on induced mammary tumor gland in Sprague-Dawley rats. Afr. J. Biotechnol., 9: 7337-7345.

4. Shukla, Y. and J. George, 2011. Combinatorial strategies employing nutraceuticals for cancer development. *Ann. N. Y. Acad. Sci.*, 1229: 162-175.
5. Wietrzyk, J., G. Gryniewicz and A. Opolski, 2005. Phytoestrogens in cancer prevention and therapy-mechanisms of their biological activity. *Anticancer Res.*, 25: 2357-2366.
6. Starley, I.F., P. Mohammed, G. Schneider and S.W. Bickler, 1999. The treatment of paediatric burns using topical papaya. *Burns*, 25: 636-639.
7. Canini, A., D. Alesiani, G. D'Arcangelo and P. Tagliatesta, 2007. Gas chromatography-mass spectrometry analysis of phenolic compounds from *Carica papaya* L. leaf. *J. Food Compos. Anal.*, 20: 584-590.
8. Zunjar, V., D. Mammen, B.M. Trivedi and M. Daniel, 2011. Pharmacognostic, physicochemical and phytochemical studies on *Carica papaya* Linn. *Pharmacogn. J.*, 3: 5-8.
9. Aravind, G., D. Bhowmik, S. Duraivel and G. Harish, 2013. Traditional and medicinal uses of *Carica papaya*. *J. Med. Plants Stud.*, 1: 7-15.
10. Otsuki, N., N.H. Dang, E. Kumagai, A. Kondo, S. Iwata and C. Morimoto, 2010. Aqueous extract of *Carica papaya* leaves exhibits anti-tumor activity and immunomodulatory effects. *J. Ethnopharmacol.*, 127: 760-767.
11. Victorio, C.P., C.L.S. Lage and R.M. Kuster, 2009. Flavonoid extraction from *Alpinia zerumbet* (Pers.) Burtt et Smith leaves using different techniques and solvents. *Eclat. Quim.*, 34: 19-24.
12. Ebada, E.S., R.A. Edrada, W. Lin and P. Proksch, 2008. Methods for isolation, purification and structural elucidation of bioactive secondary metabolites from marine invertebrates. *Nat. Protocols*, 3: 1820-1831.
13. Elmore, S., 2007. Apoptosis: A review of programmed cell death. *Toxicol. Patholol.*, 35: 495-516.
14. Althunibat, O.Y., R.B. Hashim, M. Taher, J.M. Daud, M.A. Ikeda and B.J. Zali, 2009. *In vitro* antioxidant and antiproliferative activities of three Malaysian sea cucumber species. *Eur. J. Scient. Res.*, 37: 376-387.
15. Mian, K.H. and S. Mohamed, 2001. Flavonoid (myricetin, quercetin, kaempferol, luteolin and apigenin) content of edible tropical plants. *J. Agric. Food Chem.*, 49: 3106-3112.
16. WCRF., 2000. Food, nutrition and the prevention of cancer: A global perspective. American Institute for Cancer Research, BANTA Book Group, Menasha, Wisconsin.
17. Dabrosin, C. and K. Ollinger, 1998. Protection by  $\alpha$ -tocopherol but not ascorbic acid from hydrogen peroxide induced cell death in normal human breast epithelial cells in culture. *Free Radical Res.*, 29: 227-234.
18. Yu, W., M. Simmons-Menchaca, A. Gapor, B.G. Sanders and K. Kline, 1999. Induction of apoptosis in human breast cancer cells by tocopherols and tocotrienols. *Nutr. Cancer*, 33: 26-32.
19. Malafa, M.P. and L.T. Neitzel, 2000. Vitamin E succinate promotes breast cancer tumor dormancy. *J. Surg. Res.*, 93: 163-170.
20. You, H., W. Yu, B.G. Sanders and K. Kline, 2001. *RRR- $\alpha$ -tocopheryl succinate induces MDA-MB-435 and MCF-7 human breast cancer cells to undergo differentiation. *Cell Growth Differ.*, 12: 471-480.*
21. Fleischauer, A.T., N. Simonsen and L. Arab, 2003. Antioxidant supplements and risk of breast cancer recurrence and breast cancer-related mortality among postmenopausal women. *Nutr. Cancer*, 46: 15-22.
22. Srivastava, R., R. Sharma, S. Mishra and R.B. Singh, 2011. Biochemical and molecular biological studies on oral cancer: An overview. *The Open Nutraceuticals J.*, 4: 180-188.
23. Li, Y., D. Xing, Q. Chen and W.R. Chen, 2010. Enhancement of chemotherapeutic agent-induced apoptosis by inhibition of NF- $\kappa$ B using ursolic acid. *Int. J. Cancer*, 127: 462-473.
24. Wang, X., L. Li, B. Wang and J. Xiang, 2009. Effects of ursolic acid on the proliferation and apoptosis of human ovarian cancer cells. *J. Huazhong Univ. Sci. Technol. [Med. Sci.]*, 29: 761-764.
25. Shan, J.Z., Y.Y. Xuan, S. Zheng, Q. Dong and S.Z. Zhang, 2009. Ursolic acid inhibits proliferation and induces apoptosis of HT-29 colon cancer cells by inhibiting the EGFR/MAPK pathway. *J. Zhejiang Univ. Sci. B*, 10: 668-674.
26. Manu, K.A. and G. Kuttan, 2008. Ursolic acid induces apoptosis by activating p53 and caspase-3 gene expressions and suppressing NF- $\kappa$ B mediated activation of bcl-2 in B16F-10 melanoma cells. *Int. Immunopharmacol.*, 8: 974-981.
27. Pathak, A.K., M. Bhutani, A.S. Nair, K.S. Ahn and A. Chakraborty *et al.*, 2007. Ursolic acid inhibits STAT3 activation Pathway leading to suppression of proliferation and chemosensitization of human multiple myeloma cells. *Mol. Cancer Res.*, 5: 943-955.
28. Chen, G.Q., Z.W. Yao, W.P. Zheng, L. Chen, H. Duan and Y. She, 2010. Combined antitumor effect of ursolic acid and 5-fluorouracil on human esophageal carcinoma cell Eca-109 *in vitro*. *Chin. J. Cancer Res.*, 22: 62-67.
29. Chou, C.C., J.S. Yang, H.F. Lu, S.W. Ip and C. Lo *et al.*, 2010. Quercetin-mediated cell cycle arrest and apoptosis involving activation of a caspase cascade through the mitochondrial pathway in human breast cancer MCF-7 cells. *Arch. Pharmacol. Res.*, 33: 1181-1191.
30. Obinaju, B.E. and F.L. Martin, 2010. Dose-related effects of Quercetin in the human breast carcinoma MCF-7 cell line. *Am. J. Sci. Indus. Res.*, 1: 242-261.
31. Shyu, M.H., T.C. Kao and G.C. Yen, 2010. Oleanolic acid and ursolic acid induce apoptosis in HuH7 human hepatocellular carcinoma cells through a mitochondrial-dependent pathway and downregulation of XIAP. *J. Agric. Food Chem.*, 58: 6110-6118.

32. Jeong, J.H., J.Y. An, Y.T. Kwon, J.G. Rhee and Y.J. Lee, 2009. Effects of low dose quercetin: Cancer cell-specific inhibition of cell cycle progression. *J. Cell Biochem.*, 106: 73-82.
33. Martin, R.K., 2006. Targeting apoptosis with dietary bioactive agents. *Exp. Biol. Med.*, 231: 117-129.
34. Srivastava, R., S. Akthar, R. Sharma and S. Mishra, 2015. Identification of Ellagic acid analogues as potent inhibitor of protein Kinase CK2: A chemopreventive role in oral Cancer. *Bioinformation*, 11: 21-26.
35. Mahassni, S.H. and R.M. Al-Reemi, 2013. Apoptosis and necrosis of human breast cancer cells by an aqueous extract of garden cress (*Lepidium sativum*) seeds. *Saudi J. Biol. Sci.*, 20: 131-139.
36. Ren, W., Z. Qiao, H. Wang, L. Zhu and L. Zhang, 2003. Flavonoids: Promising anticancer agents. *Med. Res. Rev.*, 23: 519-534.
37. Kuno, T., T. Tsukamoto, A. Hara and T. Tanaka, 2012. Cancer chemoprevention through the induction of apoptosis by natural compounds. *J. Biophys. Chem.*, 3: 156-173.
38. Martin, S.J. and D.R. Green, 1995. Protease activation during apoptosis: Death by a thousand cuts? *Cell*, 82: 349-352.
39. Nishida, K., O. Yamaguchi and K. Otsu, 2008. Crosstalk between autophagy and apoptosis in heart disease. *Circ. Res.*, 103: 343-351.
40. Ouyang, L., Z. Shi, S. Zhao, F.T. Wang, T.T. Zhou, B. Liu and J.K. Bao, 2012. Programmed cell death pathways in cancer: a review of apoptosis, autophagy and programmed necrosis. *Cell Proliferation*, 45: 487-498.
41. Marino, G., M. Niso-Santano, E.H. Baehrecke and G. Kroemer, 2014. Self-consumption: The interplay of autophagy and apoptosis. *Nat. Rev. Mol. Cell Biol.*, 15: 81-94.