

Chemopreventive Effects of Pomegranate Seed Oil on Skin Tumor Development in CD₁ Mice

Justin J. Hora,¹ Emily R. Maydew,¹ Ephraim P. Lansky,² and Chandradhar Dwivedi¹

¹Department of Pharmaceutical Sciences, College of Pharmacy, South Dakota State University, Brookings, SD, U.S.A.; and ²Rimonest Ltd., Horev Center, Haifa, Israel

ABSTRACT Pomegranate seed oil was investigated for possible skin cancer chemopreventive efficacy in mice. In the main experiment, two groups consisting each of 30, 4–5-week-old, female CD₁ mice were used. Both groups had skin cancer initiated with an initial topical exposure of 7,12-dimethylbenzanthracene and with biweekly promotion using 12-*O*-tetradecanoylphorbol 13-acetate (TPA). The experimental group was pretreated with 5% pomegranate seed oil prior to each TPA application. Tumor incidence, the number of mice containing at least one tumor, was 100% and 93%, and multiplicity, the average number of tumors per mouse, was 20.8 and 16.3 per mouse after 20 weeks of promotion in the control and pomegranate seed oil-treated groups, respectively ($P < .05$). In a second experiment, two groups each consisting of three CD₁ mice were used to assess the effect of pomegranate seed oil on TPA-stimulated ornithine decarboxylase (ODC) activity, an important event in skin cancer promotion. Each group received a single topical application of TPA, with the experimental group receiving a topical treatment 1 h prior with 5% pomegranate seed oil. The mice were killed 5 h later, and ODC activity was assessed by radiometric method. The experimental group showed a 17% reduction in ODC activity. Pomegranate seed oil (5%) significantly decreased ($P < .05$) tumor incidence, multiplicity, and TPA-induced ODC activity. Overall, the results highlight the potential of pomegranate seed oil as a safe and effective chemopreventive agent against skin cancer.

KEY WORDS: • ornithine decarboxylase • *Punica granatum* • initiation • promotion • prostaglandins • punicic acid

INTRODUCTION

SKIN CANCER is the most common type of cancer in the United States,¹ with more than a million reported cases² and 9,000 deaths per year.³ Increasing incidence of these cancers due to constant exposure of skin to environmental carcinogens, including both chemical agents and ultraviolet radiation, provides a strong basis for chemoprevention with both synthetic and natural, and internal and topical, remedies.⁴ Further, skin cancer chemoprevention is a useful model for cancer chemoprevention in general.⁵

Chemical and UVB radiation-induced skin carcinogenesis in murine skin and possibly human skin is a stepwise process of at least three distinct stages: initiation, promotion, and progression. Experimental initiation *in vivo* is accomplished by the topical application of a single dose of a skin carcinogen such as 7,12-dimethylbenzanthracene (DMBA), and is essentially irreversible. However, an initiation dose of carcinogen may not produce visible tumors,

resulting only following prolonged and repeated application of a tumor promoter such as 12-*O*-tetradecanoylphorbol 13-acetate (TPA) to initiated skin.^{6,7} Promoters like TPA induce ornithine decarboxylase (ODC), the rate-limiting enzyme in the synthesis of polyamines⁸ and an important molecular target for skin cancer chemoprevention.⁹ Other targets may also involve promotion, or initiation or progression events in the multistage process of neoplastic development.

Our previous work has highlighted the efficacy of topically applied natural products derived from onion and garlic oils,¹⁰ and more recently sandalwood oil^{11,12} and its constituent,¹³ in preventing skin tumors in CD₁ and SENCAR mice. In the present work, we bring this experience to bear on the study of pomegranate seed oil as a potential skin cancer chemopreventive product.

Pomegranate fruit (*Punica granatum*) has been used worldwide as an item of diet and medicine for millenia, and has also been regarded as an important symbol in world religions and mythologies and of medicine itself.¹⁴ We previously demonstrated potent antioxidant and prostaglandin-inhibitory activities for polyphenols extracted from pomegranate seed oil and pomegranate fermented juice,¹⁵ as well as a wide range of human breast cancer suppressive properties *in vitro*, including promotion of apoptosis and inhibi-

Manuscript received 5 May 2003. Revision accepted 23 May 2003.

Address reprint requests to: Chandradhar Dwivedi, Ph.D., Distinguished Professor, College of Pharmacy, Box 2202 C, South Dakota State University, Brookings, SD 57007, U.S.A., E-mail: chandradhar_dwivedi@sdstate.edu

tion of proliferation and invasion by the seed oil, and inhibition of DMBA-initiated carcinogenesis in a mouse mammary organ culture (MMOC) by the fermented juice polyphenols.¹⁶ We recently showed chemopreventive activity of the whole seed oil in the MMOC to be even stronger, weight per weight, than that of the purified fermented juice polyphenols.¹⁷

Pomegranate seed oil consists of >80% conjugated fatty acids, the most important of which is the octadecatrienoic acid, punicic acid. Punicic acid, like the <1% polyphenols in pomegranate seed oil, is an inhibitor of prostaglandin biosynthesis.¹⁸ Punicic acid is also cytotoxic to mouse leukemia cells, possibility related to inhibition of lipid peroxidation.¹⁹ Pomegranate is one of only about a half dozen plants known to contain conjugated fatty acids. A possible relationship between the relative botanical isolation of pomegranate and its singular chemistry and anticancer properties has been noted.²⁰

The purpose of the present investigation was to study the chemopreventive effects of pomegranate seed oil on DMBA-initiated and TPA-promoted skin tumor development during the initiation and promotion phases in CD₁ mice. Further, the effects of pomegranate seed oil on weight gain and ODC activity in the experimental animals were also evaluated.

MATERIALS AND METHODS

Pomegranate seed oil

Pomegranate seed oil was provided by Rimonest Ltd. (Rimonest Ltd., Haifa, Israel; www.rimonest.com) from pomegranates of the "Wonderful" cultivar, organically grown at Kibbutz Sde Eliahu, Israel, in the year 2000. Seeds were separated from their juice sacs, washed in water, and dried in a convection current solar dryer. Oil extrusion was by "cold press" at 80°C, using a Type 40A electric screw press (Skeppsta Maskin, Orebro, Sweden). The oil was assayed by an independent laboratory (Mylnfield Research Services, Invergowrie, Dundee, Scotland) and shown to contain not less than 80% conjugated fatty acids as triglycerols, diglycerols, and monoglycerols.

Tumorogenesis protocol

The skin cancer protocol of Dwivedi *et al.*¹³ was used. In brief, 4–6-week-old CD₁ mice were divided into two groups, each group containing 30 mice, as indicated in Table 1. The mice were kept in an environmentally controlled room with temperature, humidity, and light regulated. The backs of the

mice were shaved carefully with an electric clipper to avoid cuts. The mice were allowed to rest for 2 days before carcinogenesis was initiated.

Carcinogenesis was initiated with DMBA (200 nmol in 100 μ L of acetone) applied topically. One week later, carcinogenesis was promoted with TPA (5 nmol in 100 μ L of acetone), applied topically twice weekly. TPA treatment continued throughout the duration of the experiment (20 weeks). Mice in group 1 served as the control and were pretreated topically with 100 μ L of acetone 1 h prior to each TPA application. Mice in group 2 were pretreated topically with 100 μ L of 5% pomegranate seed oil in acetone 1 h prior to each TPA application. Tumor counts and group weights were taken on a weekly basis. Tumor incidence and multiplicity were calculated and analyzed statistically.

ODC assay

Mice were divided into two groups, each containing three mice. The backs of the mice were shaved carefully with an electric clipper to avoid cuts. Mice in group 1 received 100 μ L of acetone before TPA (5 nmol in 100 μ L of acetone) treatment topically. Mice in group 2 received 100 μ L of 5% pomegranate seed oil in acetone, before topical TPA (5 nmol in 100 μ L of acetone) treatment.

Mice were killed 5 h after the topical applications of TPA. The dorsal epidermis was removed and homogenized in phosphate buffer (pH 7.2) containing 0.1 mM pyridoxal phosphate and 0.1 mM EDTA. The homogenate was centrifuged at 105,000 *g* for 90 min and the supernatant collected and used for the ODC assay. The assay mixture in the main part of a Warburg flask was composed of 40 μ L of phosphate buffer (pH 7.2), 25 μ L of pyridoxal phosphate, 25 μ L of dithiothreitol, 25 μ L of EDTA, 10 μ L of L-ornithine containing 0.5 μ Ci of DL-[1-¹⁴C]ornithine, and 200 μ L of epidermal supernatant.

The center well of the Warburg flask contained 400 μ L of ethanolamine and methoxyethanol used to absorb the ¹⁴CO₂ produced in the main compartment. After incubation at 37°C for 1 h, the reaction was stopped by the addition of 500 μ L of citric acid. The mixture was stored in a dark place overnight to ensure complete absorption of ¹⁴CO₂ in the center well. The contents of the center well were transferred to a scintillation vial. The center well was washed with 0.5 mL of ethanol four times, and the wash also added to the scintillation vial, along with 10 mL of scintillation fluor. Radioactivity was counted with a Beckman LS6000SE liquid scintillation counter. The disintegrations per minute were quantified. Assessment of ODC activity was accomplished by measuring the production of ¹⁴CO₂ from DL-[1-¹⁴C]ornithine.

Protein assay

Protein was assayed in the supernatant with a Bio-Rad Protein Assay Kit. A standard curve was obtained using bovine serum albumin. Absorbance values at 595 nm were determined using the spectrophotometer. Protein concentra-

TABLE 1. TOPICAL TREATMENTS RECEIVED BY THE MICE IN THE TWO EXPERIMENTS

Group 1	100 μ L of acetone
Group 2	100 μ L of 5% pomegranate seed oil in acetone

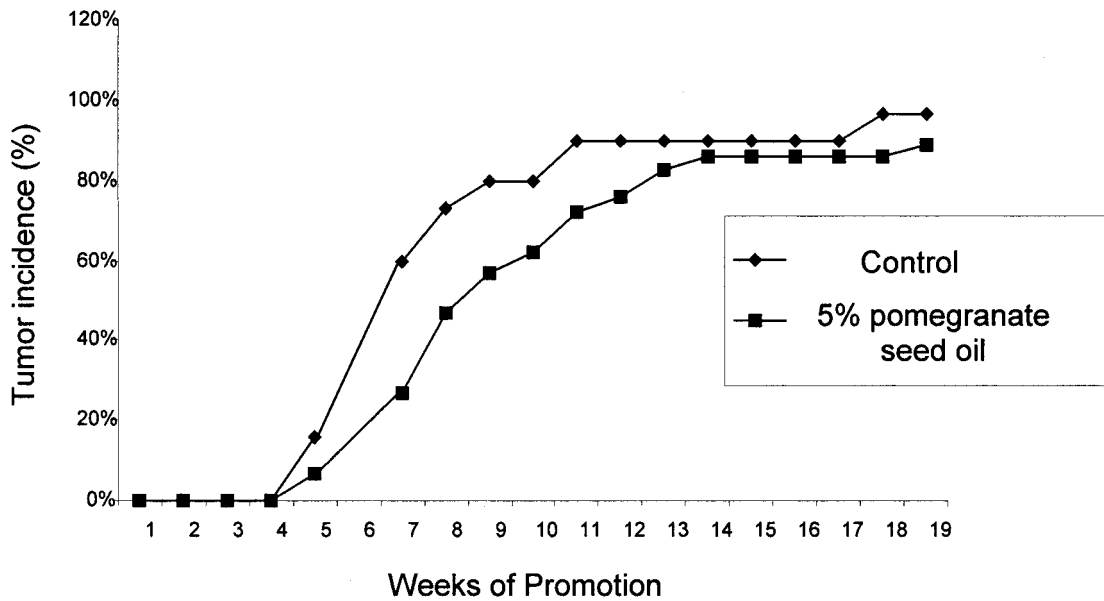


FIG. 1. The effects of pomegranate seed oil treatment on tumor incidence in CD₁ mice.

tions of the supernatant were extrapolated from the standard curve data.

Statistical analysis

The INSTAT software (GraphPad, San Diego, CA, U.S.A.) was used for the data analysis. χ^2 was used for the comparison of papilloma incidence and Student's *t* test for tumor multiplicity and ODC activity. Significance was considered at $P < .05$.

RESULTS

The effects of pomegranate seed oil treatment on the incidence of skin tumors in CD₁ mice are shown in Fig. 1. Skin tumors appeared in the sixth week of promotion after the initial DMBA application in the control and treated groups. Pomegranate seed oil treatment did not delay the appearance of tumors, but significantly decreased ($P < .05$) the rate at which the tumors developed. Skin tumor incidence after 20 weeks of promotion was 100% and 93% for

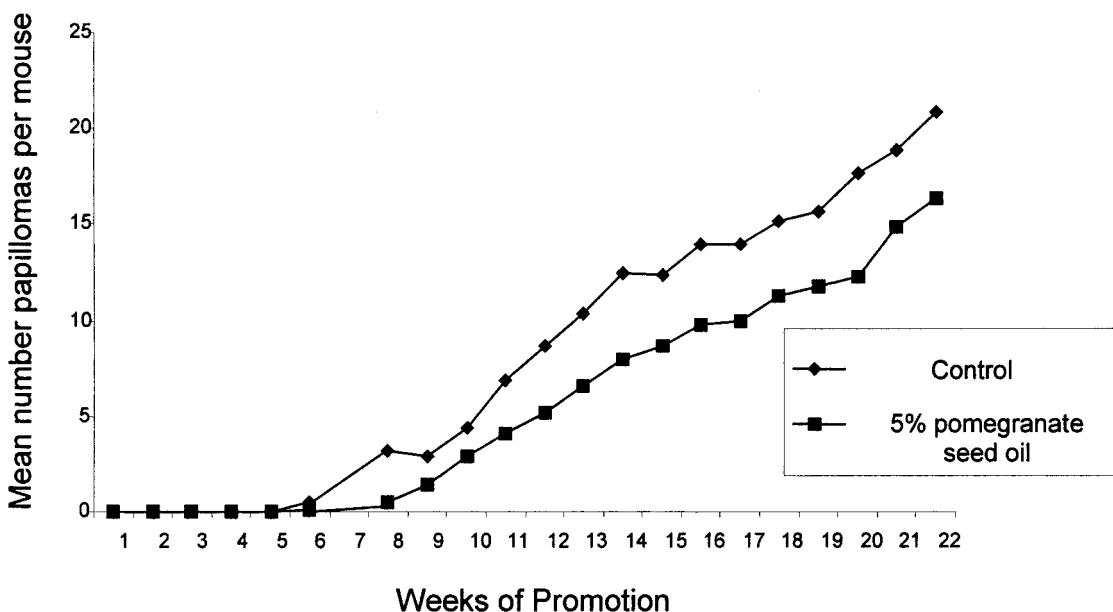


FIG. 2. The effects of pomegranate seed oil treatment on tumor multiplicity in CD₁ mice.

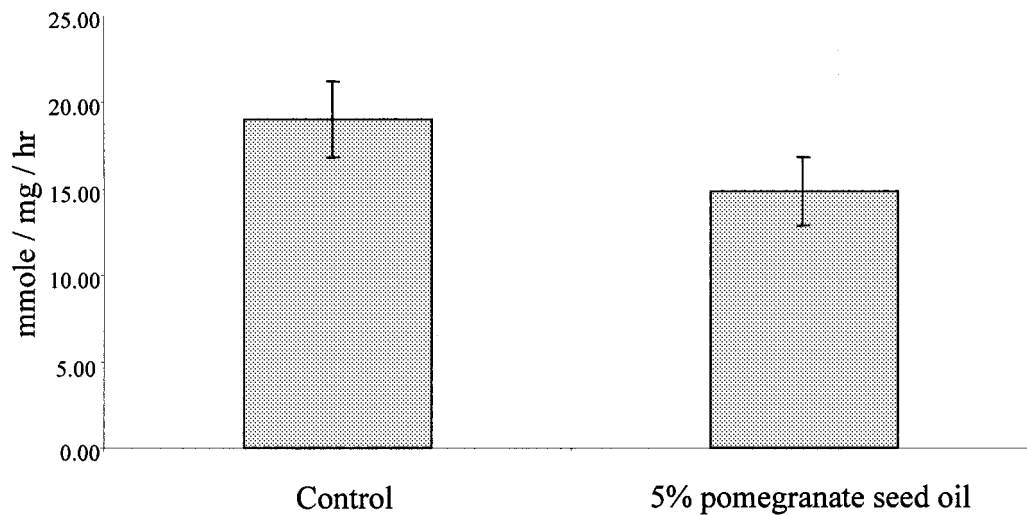


FIG. 3. The effects of pomegranate oil treatment on TPA-induced epidermal ODC activity in CD₁ mice. ODC activity is expressed as nmol of ¹⁴CO₂ produced/ μ g of protein/h. The 5% pomegranate seed oil (only) group did not have detectable ODC activity (data not shown).

the control and 5% pomegranate seed oil-treated groups, respectively.

The effects of pomegranate seed oil treatment on tumor multiplicity in CD₁ mice are shown in Fig. 2. Pomegranate seed oil treatment significantly decreased ($P < .05$) the tumor multiplicity throughout the 20 weeks of promotion. The mean number of tumors per mouse was 20.8 and 16.3 for the control and 5% pomegranate seed oil-treated groups, respectively.

Topical application of 5% pomegranate seed oil also significantly inhibited ($P < .05$) TPA-induced epidermal ODC activity. Fig. 3 illustrates the effects of pomegranate seed oil treatment on TPA-induced epidermal ODC activity. The

ODC activity was 18.49 and 14.84 nmol of ¹⁴CO₂/mg/h in the control and 5% pomegranate seed oil-treated groups, respectively. The pomegranate seed oil group has significantly ($P < .05$) decreased ODC activity. Topical application of 5% pomegranate seed oil alone did not induce any epidermal ODC activity. Topical application of 5% pomegranate seed oil also did not have any effect on weight gain, as indicated in Fig. 4.

CONCLUSIONS

Pomegranate seed oil (5%) topical applications significantly decreased the incidence of skin tumor development,

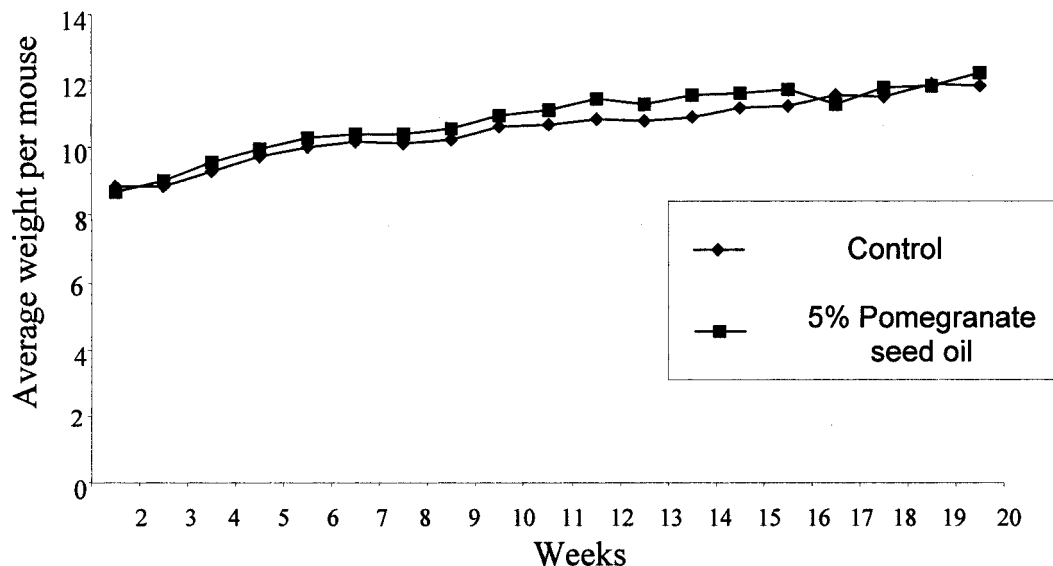


FIG. 4. The effects of pomegranate seed oil treatment on weight gain.

skin tumor multiplicity, and ornithine decarboxylase activity during 20 weeks of promotion. It is thus likely that the inhibition of ornithine decarboxylase by the pomegranate seed oil was at least partially responsible for the chemopreventive effect.

As noted, pomegranate seed oil is very rich in punicic acid, a known inhibitor of prostaglandin biosynthesis, specifically by inhibiting cyclooxygenase (Cox 1 and Cox 2) and lipoxygenase.²¹ Pomegranate seed oil also inhibits the upstream eicosanoid enzyme, phospholipase A2, expressed by human prostate cancer cells.²² That prostaglandins at very low concentrations promote ornithine decarboxylase²³ suggests that the inhibition of prostaglandin biosynthesis by pomegranate seed oil might also contribute to its inhibition of ornithine decarboxylase and, ultimately, to inhibition of skin cancer promotion.

Overall, pomegranate seed oil appears to be a benign natural product with potential as a topical chemopreventive agent against skin cancer. More in-depth investigations, including clinical studies, are warranted to evaluate this hypothesis further.

ACKNOWLEDGMENTS

The authors wish to thank Mr. Eli Merom of Kibbutz Sde Eliahu, Israel, for supplying the organically grown pomegranates used in this study. Thanks also to Alexander Botvinnik for technical assistance in the preparation of the manuscript.

REFERENCES

1. Skin cancer (what you need to know about). National Institutes of Health, National Cancer Institute, NIH Publication no. 94-1563 (Revised April 1993).
2. Facts on Skin Cancer. American Cancer Society Publication 99-200M. Rev. 8/95, no. 2049.
3. Cancer Facts & Figures—1996. American Cancer Society Publication 96-300M, no. 5008-96, p. 17.
4. Gupta S, Mukhtar H: Chemoprevention of skin cancer: current status and future prospects. *Cancer Metastasis Rev* 2002;21:363–380.
5. Richmond E, Viner JL: Chemoprevention of skin cancer. *Semin Oncol Nurs* 2003;19:62–69.
6. Agarwal R, Mukhtar H: Cutaneous chemical carcinogens. In: *Pharmacology of the Skin* (Mukhtar H, ed.). CRC Press, Boca Raton, FL, 1991, pp. 371–387.
7. Boutwell RK: Some biological aspects of skin carcinogenesis. *Prog Exp Tumor Res* 1984;4:207–250.
8. O'Brien TG, Simsiman RC, Boutwell RK. Induction of the polyamine-biosynthetic enzymes in mouse epidermis by tumor-promoting agents. *Cancer Res* 1975;35:1662–1670.
9. Stratton SP, Dorr RT, Alberts DS. The state-of-the-art in chemoprevention of skin cancer. *Eur J Cancer* 2000;36:1292–1297.
10. Dwivedi C, Rohlf S, Jarvis D, Engineer FN. Chemoprevention of chemically induced skin tumor development by diallyl sulfide and diallyl disulfide. *Pharm Res* 1992;9:1668–1670.
11. Dwivedi C, Abu-Ghazaleh A. Chemopreventive effects of sandalwood oil on skin papillomas in mice. *Eur J Cancer Prev* 1997;6:399–401.
12. Dwivedi C, Zhang Y: Sandalwood oil prevents skin tumour development in CD1 mice. *Eur J Cancer Prev* 1999;8:449–455.
13. Dwivedi C, Guan X, Harmsen WL, Voss AL, Goetz-Parten D, Koopman EM, Johnson KM, Valluri HB, Matthees DD: Chemopreventive effects of α -santalol on skin tumor development in CD₁ and Sencar mice. *Cancer Epidemiol Biomarkers Prev* 2003;12:151–156.
14. Langley P. Why a pomegranate? *BMJ* 2000;321:1153–1154.
15. Schubert SY, Lansky EP, Neeman I. Antioxidant and eicosanoid enzyme inhibition properties of pomegranate seed oil and fermented juice flavonoids. *J Ethnopharmacol* 1999;66:11–17.
16. Kim ND, Mehta R, Yu W, Neeman I, Livney T, Amichay A, Poirier D, Nicholls P, Kirby A, Jiang W, Mansel R, Ramachandran C, Rabi T, Kaplan B, Lansky E. Chemopreventive and adjuvant therapeutic potential of pomegranate (*Punica granatum*) for human breast cancer. *Breast Cancer Res Treat* 2002;71:203–217.
17. Unpublished data presented at the 13th Annual Meeting of the North American Menopause Society, Chicago 2002. Rajendra Mehta, Department of Surgical Oncology, University of Illinois at Chicago.
18. Nugteren DH, Christ-Hazelhof E. Naturally occurring conjugated octadecatrienoic acids are strong inhibitors of prostaglandin biosynthesis. *Prostaglandins* 1987;33:403–417.
19. Suzuki R, Noguchi R, Ota T, Abe M, Miyashita K, Kawada T: Cytotoxic effect of conjugated trienoic fatty acids on mouse tumor and human monocytic leukemia cells. *Lipids* 2001;36:477–482.
20. Longtin R: The pomegranate: nature's power fruit? *J Natl Cancer Inst* 2003;95:346–348.
21. Unpublished data, Robert Newman, Department of Pharmacology, MD Anderson Cancer Center, Houston, TX.
22. Unpublished data, Wenguo Jiang, Department of Surgery, Cardiff University, U.K.
23. Cameron CM, Rillema JA: Effects of prostaglandins on the prolactin stimulation of lipid biosynthesis in mouse mammary gland explants. *Prostaglandins Leukot Med* 1983;10:433–441.